

Kenyon College

Digital Kenyon: Research, Scholarship, and Creative Exchange

Faculty Publications

Biochemistry and Molecular Biology

2005

pH Regulates Genes for Flagellar Motility, Catabolism, and Oxidative Stress in *Escherichia coli* K-12†

Joan Slonczewski

Kenyon College, slonczewski@kenyon.edu

Follow this and additional works at: https://digital.kenyon.edu/biochem_publications



Part of the [Biochemistry, Biophysics, and Structural Biology Commons](#), and the [Biology Commons](#)

Recommended Citation

Maurer, L. M.,* E. Yohannes, S. S. BonDurant, M. Radmacher, and J. L. Slonczewski. 2005. pH regulates genes for flagellar motility, catabolism, and oxidative stress in *Escherichia coli* K-12. *J Bacteriol.* 187:304-319

This Article is brought to you for free and open access by the Biochemistry and Molecular Biology at Digital Kenyon: Research, Scholarship, and Creative Exchange. It has been accepted for inclusion in Faculty Publications by an authorized administrator of Digital Kenyon: Research, Scholarship, and Creative Exchange. For more information, please contact noltj@kenyon.edu.

pH Regulates Genes for Flagellar Motility, Catabolism, and Oxidative Stress in *Escherichia coli* K-12†

Lisa M. Maurer,¹ Elizabeth Yohannes,¹ Sandra S. Bondurant,² Michael Radmacher,¹
and Joan L. Slonczewski^{1*}

Department of Biology, Kenyon College, Gambier, Ohio,¹ and Gene Expression Center,
University of Wisconsin, Madison, Wisconsin²

Received 30 July 2004/Accepted 17 September 2004

Gene expression profiles of *Escherichia coli* K-12 W3110 were compared as a function of steady-state external pH. Cultures were grown to an optical density at 600 nm of 0.3 in potassium-modified Luria-Bertani medium buffered at pH 5.0, 7.0, and 8.7. For each of the three pH conditions, cDNA from RNA of five independent cultures was hybridized to Affymetrix *E. coli* arrays. Analysis of variance with an α level of 0.001 resulted in 98% power to detect genes showing a twofold difference in expression. Normalized expression indices were calculated for each gene and intergenic region (IG). Differential expression among the three pH classes was observed for 763 genes and 353 IGs. Hierarchical clustering yielded six well-defined clusters of pH profiles, designated Acid High (highest expression at pH 5.0), Acid Low (lowest expression at pH 5.0), Base High (highest at pH 8.7), Base Low (lowest at pH 8.7), Neutral High (highest at pH 7.0, lower in acid or base), and Neutral Low (lowest at pH 7.0, higher at both pH extremes). Flagellar and chemotaxis genes were repressed at pH 8.7 (Base Low cluster), where the cell's transmembrane proton potential is diminished by the maintenance of an inverted pH gradient. High pH also repressed the proton pumps cytochrome *o* (*cyo*) and NADH dehydrogenases I and II. By contrast, the proton-importing ATP synthase F_1F_0 and the microaerophilic cytochrome *d* (*cyd*), which minimizes proton export, were induced at pH 8.7. These observations are consistent with a model in which high pH represses synthesis of flagella, which expend proton motive force, while stepping up electron transport and ATPase components that keep protons inside the cell. Acid-induced genes, on the other hand, were coinduced by conditions associated with increased metabolic rate, such as oxidative stress. All six pH-dependent clusters included envelope and periplasmic proteins, which directly experience external pH. Overall, this study showed that (i) low pH accelerates acid consumption and proton export, while coinducing oxidative stress and heat shock regulons; (ii) high pH accelerates proton import, while repressing the energy-expensive flagellar and chemotaxis regulons; and (iii) pH differentially regulates a large number of periplasmic and envelope proteins.

Escherichia coli and related enteric bacteria respond to a wide range of pH stresses by regulating gene expression (for reviews see references 21 and 68) and protein profiles (73, 82). Enteric bacteria encounter a wide range of external pHs in their natural habitat, the human digestive tract (17). Colonization of the intestine requires transient survival through the stomach at pH 1 to 2 (fasting) or 2 to 7 (transiently, during feeding) (18), as well as exposure to pancreatic secretions at pH 10 (25) followed by growth and persistence at a range of external pHs of 5 to 8 (20). Growth at a pH substantially higher or lower than the cytoplasmic pH 7.6 induces protective responses with two fundamental aims: to maintain internal pH homeostasis and to prepare the cell to survive future exposure to more extreme pH conditions (below pH 5 or above pH 9) that no longer permit growth (11, 41, 70).

The effects of pH on enteric bacteria contribute to disease. Low pH enhances expression of numerous virulence factors, such as the ToxR-ToxT virulence regulon in *Vibrio cholerae* (7), the *phoP-phoQ* regulon of *Salmonella enterica* (6), and the pH 6 antigen of *Yersinia pestis* (50). Acid stress contributes to

food preservation; many food preservatives are membrane-permeant acids whose uptake is enhanced by acid (60), and acid interacts in complex ways with both temperature and organic food preservatives (65).

While growth in acid challenges pH homeostasis, the pH difference across the inner cell membrane (Δ pH) nevertheless contributes cell energy in the form of proton potential or proton motive force (Δ p). The proton potential powers motility, ATP synthesis, and catabolite transport (for a review see reference 29). But low pH also amplifies the uptake of membrane-permeant acids that dissipate the proton potential (59). Thus, we expect low pH to induce a combination of positive and negative responses.

Much of bacterial catabolism affects pH, and in *E. coli* a growing number of catabolic enzymes and catabolite transporters are known to be regulated by pH (21, 73). Sugar fermentation initially generates short-chain acids that are excreted but accumulate and reenter the cytoplasm, causing acidification. Thus, it is not surprising that sugar transporters such as OmpF and the maltose regulon are down-regulated at low pH (13). Consumption of acids by the tricarboxylic acid (TCA) cycle causes alkalization, a common result of growth to stationary phase in tryptone-based media (66, 73). Catabolism of amino acids by decarboxylases generates alkaline amines, which help the cell counteract external acidification, for example, the lysine and arginine decarboxylases (4, 27, 45, 47, 71). High pH,

* Corresponding author. Mailing address: Department of Biology, Higley Hall, 202 N. College Dr., Kenyon College, Gambier, OH 43022. Phone: (740) 427-5397. Fax: (740) 427-5089. E-mail: slonczewski@kenyon.edu.

† Supplemental material for this article may be found at <http://jb.asm.org/>.

however, induces deaminases that generate acids, such as tryptophan deaminase (*tnaAB*) and serine deaminase (*sda*) (9, 73, 82).

A complicated case is that of the glutamic acid decarboxylase genes *gadA* and *gadBC* (12, 44). The *gad* system enables cells to survive extreme acid (77), but its expression is induced mainly at high pH, or in Luria-Bertani medium grown to stationary phase, where pH naturally increases (73, 82). An alternative role of *gad*, particularly under anaerobiosis, may be to channel its product γ -aminobutyric acid into fermentation acids.

Even mild acid (pH 6 to 7) greatly amplifies the uptake of membrane-permeant weak acids such as acetate. Permeant acids pass through the bacterial membrane and dissociate in the cytoplasm, causing accumulation of anions and depression of internal pH (34, 56). Acetate concentrations rise as cell density increases, and acetate induces a large number of genes and proteins (3, 35). Growth inhibition occurs as a result of both lower internal pH and the differential ability of anions to inhibit metabolism (60). The effect of permeant acids is critical in the human colon, where the concentration of short-chain fatty acids totals approximately 100 mM (15).

While numerous responses to pH stress are known, the mechanisms by which *E. coli* maintains its internal pH at 7.6 remain poorly understood. The electron transport chain pumps protons outside the cell, and the H^+ -ATPase either exports or imports protons, but mutants in these components maintain pH homeostasis. There is evidence that potassium exchange contributes to pH homeostasis in external acid (5, 10, 52, 80), but the precise mechanisms remain unclear. At high pH, the electrical potential ($\Delta\psi$) is diminished in order to compensate for the inverted ΔpH . The sodium-proton antiporter NhaA contributes to internal pH maintenance under sodium stress (24, 75). High pH also induces major stress systems such as heat shock response (1, 28, 74), the SOS regulon (63), and the CpxP envelope stress response (16).

At more extreme pH values, well below the growth range (as low as pH 1.5 for clinical isolates) *E. coli* can retain viability for many hours, a phenomenon termed acid survival or acid resistance. Acid resistance is enhanced by many genes induced during growth at the acid end of the pH range (pH 5) or growth to stationary phase. Acid-induced acid resistance factors include periplasmic chaperones such as the *hdeA* product (23), envelope proteins such as OsmY, and redox modulators such as Tpx (73, 78). A complex acid resistance regulon including the *gad* system is regulated by transcription factors GadX-GadW and EvgA-YdeO, as well as by RpoS, H-NS, and cyclic AMP (11, 12, 44, 79). *E. coli* also exhibits base resistance, the ability to survive at or above pH 10 (58, 70). Base resistance requires *rpoS* and components of the *gad* system (30).

Finally, pH may affect flagellar motility, although the present picture is unclear. According to one report, growth in acid represses flagellar genes and eliminates motility (72), whereas another group finds motility enhanced by acetate and propionate, which cause acid stress (53).

To investigate acid and base response, we used microarrays to compare *E. coli* gene expression at low, neutral, or high external pH. Past microarray studies of pH response have been limited by their absence of pH conditions above pH 7 (44, 78); their use of glucose minimal medium (78), in which many

catabolic genes are repressed; and their focus on only a single acid resistance regulon (44). Our experimental design included both acid and base conditions, as well as pH 7.0. For each growth condition, five independent cultures were hybridized separately, a number of replicates that ensured detection of virtually all expression ratios of at least twofold. The coregulation of numerous genes within operons confirmed the biological relevance of our expression ratios. Our study revealed unexpected patterns of pH response and clarified the overlap of pH stress with other stress responses.

MATERIALS AND METHODS

Growth conditions. *E. coli* K-12 strain W3110 (R. VanBogelen and F. Neidhardt) was grown overnight in unbuffered potassium-modified Luria broth (LBK) (10 g of tryptone/liter, 5 g of yeast extract/liter, 7.45 g of KCl/liter). For pH-controlled growth, media were buffered with 100 mM homopiperazine-*N,N'*-bis-2-(ethanesulfonic acid) (HOMOPIPES) (pK_a , 4.55 and 8.12). The pH of the media were adjusted to 5.0, 7.0, or 8.7 with KOH solution to avoid extra sodium ions, which stress cells at high pH (24). To maximize aeration and maintain logarithmic growth, the overnight culture was diluted 1,000-fold into 12 ml of buffered medium in a 125-ml baffled flask and rotated at 240 rpm. Cultures were grown at 37°C to an optical density at 600 nm of 0.3. For all cultures, the pH was tested after growth to ensure that the values were maintained at ± 0.2 pH unit of the pH of the original uninoculated medium.

To observe motility, we used *E. coli* K-12 strain RP437 and *S. enterica* serovar Typhimurium SJW1103 from a laboratory in which strains are maintained for motility (M. Macnab). Culture was spotted on tryptone-KCl soft-agar plates (0.35% Bacto Agar) and incubated at 37°C until cells swam out. Culture was picked from the leading edge of the swimming cells and inoculated into LBK for overnight growth. For quantitative assay of motility, 5 μ l of culture was spotted in triplicate on plates containing tryptone-KCl with 100 mM sulfonate buffer of appropriate pK_a (73). After growth for 8 h, the diameter of motile cell growth was measured.

RNA isolation. Bacterial RNA was isolated using the Qiagen RNeasy kit with on-column DNA digestion (Qiagen), with additional DNA removal with Ambion DNase. To perform this additional DNase digestion, RNA was precipitated and redissolved in 85 μ l of nuclease-free water. We then added 10 μ l of 10 \times DNase I buffer and 5 μ l of (1-U/ μ l) DNase I (Ambion). The DNase reaction mixture was incubated at 37°C for 30 min and then chilled on ice. A second RNeasy column purification was performed.

cDNA preparation and array hybridization. For microarrays, standard methods were used for cDNA synthesis, fragmentation, and end-terminus biotin labeling, based on Affymetrix protocols. Labeled cDNA was hybridized to *E. coli* Affymetrix Antisense Genome Arrays. Hybridized arrays were stained with streptavidin-phycoerythrin with the use of the Affymetrix Fluidic Station. After staining, arrays were scanned with a GC2500 scanner.

Statistical analysis of gene expression. The experiment was designed so as to minimize both false-positive and false-negative results for expressed genes. Five full replicates (with respect to *E. coli* growth, RNA isolation, sample preparation, and array hybridization) were performed for each pH condition.

The median within-group variance in expression for all genes in the data set was 0.031 (or standard deviation, 0.175). To test for significant differences in expression between the pH classes, one-way analysis of variance (ANOVA) was performed at a significance level of 0.001; thus, of every thousand genes tested, only one false positive would be expected. For a gene with average within-group variability, our sample size provided statistical power of 98% to detect a twofold difference in gene expression among pH groups. That is, only 2% of genes that show a twofold difference in expression between any two pH groups would be missed (false negatives).

Model-based expression analysis with dChip software (40) was performed on the probe-level data from Affymetrix's DAT files. The model relates target RNA levels to the probe signals by a linear function that weights the significance of all oligonucleotide probes for each gene. The analysis includes normalization, which rescales data from different arrays so that comparisons can be made among arrays. Each array was normalized to a baseline array from a pH 7 culture, by using local regression on an invariant set of probes (62). Model-based expression indices were calculated for each gene on each array by using only the perfect match probes (61), and outlier detection was performed (39). Only probe sets that received an Affymetrix call of "present" on greater than 50% of the arrays were used in subsequent analyses. "Present" or "absent" calls use information

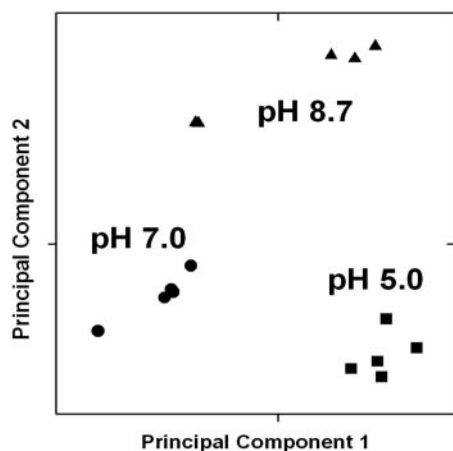


FIG. 1. Principal component analysis. The gene expression profiles of the arrays were visualized in two-dimensional Euclidian space, by using BRB ArrayTools software as described under Materials and Methods. The first and second principal components are shown. pH 5.0, squares; pH 7.0, circles; pH 8.7, triangles.

from paired perfect-match and single-base-mismatch probes. Four thousand six hundred fifty probe sets passed this criterion.

For genes whose probe sets passed the 50% screen, one-way ANOVA was performed on the \log_2 -transformed model-based expression indices, on a gene-by-gene basis. For each gene that displayed significant differences in expression among the classes, pairwise comparisons of pH classes were determined using Tukey's multiple comparisons procedure to control the familywise error rate for the t test.

Additional analyses were performed to explore categories of differential gene expression. Global relationships among arrays were visualized by performing a principal component analysis (81) on the expression data and plotting arrays in two-dimensional space corresponding to the first two principal components. The gene expression profiles of the arrays were visualized in two-dimensional Euclidian space, by using BRB ArrayTools software. In addition, categories of differential expression profiles across the pH classes were generated by a hierarchical cluster analysis of differentially expressed genes, based on the average linkage method (19) with BRB ArrayTools.

RESULTS

Growth range of pH. To study the full range of pH response, we selected the widest pH range (pH 5.0 to 8.7) in which cultures maintained reasonable doubling times and approximately constant pH throughout growth. Culture media were

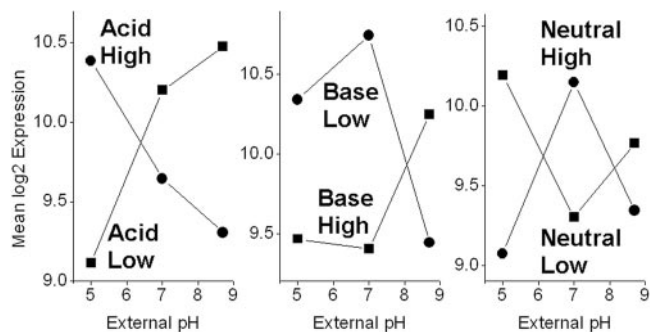


FIG. 3. Cluster mean expression profiles. The mean expression profiles over pH are plotted for the six clusters defined in Fig. 2.

adjusted to pH 5.0, 7.0, and 8.7. The doubling time for *E. coli* cultured at pH 5.0 and 8.7 was approximately 25 min and at pH 7.0 was 18 min. All cultures were grown to an optical density of 0.3 in order to facilitate at least five complete replications. The final pH of growth cultures was found to be within ± 0.2 of the initial pH. The internal pH of the cytoplasm is approximately 7.6 (69); thus, growth at external pH 7.0 might induce some acid response.

Probe hybridization. To determine differential gene expression, the \log_2 transforms of normalized model-based expression values of genes were compared. Of the 7,231 genes and intergenic regions (IGs) on the array, 4,650 loci were detected on more than half (eight or more) of the 15 arrays. These loci, constituting about 70% of the total array, were taken for further analysis.

Principal component analysis. Global relationships among arrays were visualized by performing a principal component analysis (81) on the expression data (Fig. 1). Before dimensional reduction, each array existed in 4,650-dimensional space (one dimension for each of the 4,650 intensity values). The array comparisons were plotted in two-dimensional space, corresponding to the first and second principal components of variation. The first principal component for each array is the weighted linear combination of intensity values that shows maximum variation, whereas the second principal component is a weighted linear combination orthogonal to the first component that has maximum variance.



FIG. 2. Cluster analysis of differentially expressed genes. The dendrogram was generated based on the average linkage method (19) with BRB ArrayTools. At a correlation of 0.6, six clusters of related gene expression were designated Acid High (AH), Acid Low (AL), Base High (BH), Base Low (BL), Neutral High (NH), and Neutral Low (NL).

The principal component analysis indicated that the microarrays from each of the three pH conditions appeared in distinct groups (Fig. 1). Within-class variability was small relative to variability among pH levels. The pH 8.7 arrays showed the greatest degree of separation, clustering into two groups based on the date on which the arrays were hybridized, but this difference was small compared to the differences between pH classes.

ANOVA for significance of expression profiles. We compared gene expression among the three pH groups on a gene-by-gene basis using one-way ANOVA at a significance level of 0.001. The significance level indicates the probability of a false positive, and we therefore expect $0.001 \times 4,650 = 4.65$ false-positive genes (i.e., genes that are not truly differentially expressed but that appear in our differentially expressed list) in our full analysis. Of the 4,650 loci with eight or more “present” calls on arrays, 761 genes and 353 IGs showed a significant F value for differential expression among the three pH classes. Thus, about 17% of *E. coli* genes showed significant modulation of expression as a function of pH.

Cluster analysis. As a first attempt at categorizing differentially expressed genes, we performed a hierarchical cluster analysis (19) of differentially expressed genes (Fig. 2). We used average linkage and one minus the centered Pearson correlation as the distance metric. At a correlation value of approximately 0.6, the dendrogram generated six clusters of gene expression profiles.

Within each of the six clusters, the average profiles were determined for all the gene expression indices (\log_2 intensity values) across the three pH conditions (Fig. 3). The clusters were defined by their mean expression profiles across the three pH conditions. The Acid High cluster showed highest expression at pH 5.0, declining at pH 7.0 and 8.7. It included 160 genes and 49 IGs. Acid Low (113 genes, 57 IGs) showed approximately the reverse profile, with its lowest expression at pH 5.0, rising at pH 7.0 and 8.7. Base High (93 genes, 70 IGs) showed low expression at pH 5.0 and 7.0 and higher expression at pH 8.7, whereas Base Low (123 genes, 40 IGs) showed the reverse, higher expression at pH 5.0 and 7.0 than at pH 8.7. The Neutral High cluster (93 genes, 14 IGs) showed highest expression at pH 7.0 and lower expression at both pH extremes. The Neutral Low cluster (181 genes, 123 IGs) showed the lowest expression at pH 7.0 and higher expression at both pH extremes, although the mean expression was substantially greater at pH 5.0 than at pH 8.7; a number of acid-induced genes fell in this category.

Table 1 lists the genes that fell into each cluster; details of description and Blattner open reading frame (ORF) numbers are available online in Table S1 in the supplemental material. In many cases, all or most of the ORFs in a given operon were induced in the same cluster; see, for example, the *atp* operon (Base High cluster) and the *flg* and *fli* operons (Base Low cluster).

Known acid-induced genes and acid resistance genes such as *sucBC* and *hdeA* (73) generally fell under Acid High, Base Low, or Neutral Low, a cluster whose mean expression indices were actually twofold higher in acid than in base (Fig. 3). These results are generally consistent with the cluster pH profiles and with the structure of the cluster dendrogram, in which the Acid High profile correlates most closely with the Neutral Low pro-

file. Most known base-induced genes, such as *alk* (*ygjT*) (8, 73) and *tnaA* (9), fell under Base High or Acid Low.

For IGs, the cluster assignment and expression ratios are presented online in Table S2 in the supplemental material. Expression of an IG may result from a small regulatory RNA that lies between protein-encoding genes (2, 43), or it may indicate the tail end of mRNA containing pH-regulated genes. For example, the IGs upstream of *tnaC* (*tnaA* leader peptide) and downstream of *tnaB* both were repressed in acid, as are *tnaA* and *tnaB*.

Individual gene expression ratios. For genes whose overall expression profile yielded a significant F value (one-way ANOVA), we used the Tukey procedure to determine ratios of average model-based expression indices from cultures at pH 5.0 versus pH 7.0, at pH 8.7 versus pH 7.0, and at pH 8.7 versus pH 5.0. The full list of individual \log_2 expression ratios for all analyzed genes is presented in Table S1 in the supplemental material and for IGs is presented in Table S2 in the supplemental material; for genes of particular interest grouped in functional categories, the data are presented in Tables 3 through 7. Expression ratios that are significant at $\alpha = 0.001$ are shown in boldface.

The genes most strongly regulated by pH are summarized in Table 2. These genes each showed an expression ratio of at least fourfold ($\log_2 = 2$) between two of the pH classes. Note that the two genes most strongly induced in acid are ORFs with no known function, *yhcN* and *yagU*. Other acid-induced genes include those for catabolic enzymes in pathways that consume acids, such as *sdhCD* (succinate dehydrogenase). Genes repressed at high pH include several members of the flagellar regulon, including the main flagellar subunit *fliC* (for a review see reference 42).

The genes most strongly induced at high pH included *tnaC*, encoding the tryptophanase leader peptide (26), as well as *tnaA* (tryptophanase) and the Trp transporter gene *tnaB*, with its leader peptide gene *tnaC*. Previously in proteomic gels, we found tryptophanase to be the most highly expressed protein observed at high pH (9). The alkali-inducible protease gene *cpxP* (16) was also strongly induced. Members of the maltose transport regulon (*malEKM*) were strongly repressed by acid, consistent with previous reports (31, 73). But proteins strongly induced by base also included those from genes of unknown function, such as *yifO* and *ymgD*.

Flagellar and chemotaxis regulons. Motility in *E. coli* is governed by the flagellar chemotaxis regulon including 50 components in 19 operons, governed by the major regulators FlhC and FlhD (42, 76). The expression of the regulatory operon *flhCD* is controlled by numerous environmental response systems, such as adenylate cyclase (37), RcsCDB (22), and ClpXP (76).

Nearly all the genes of the flagellar regulons (47 genes) were repressed at high pH (Table 2). Forty-one genes fell in the Base Low cluster, which means that the bulk of significant expression difference occurred between pH 7.0 and 8.7. (The other six genes were Acid High.) These genes were among the most strongly base-repressed genes in the arrays (Table 2); for instance, *fliC*, encoding the flagellin monomer, had the lowest pH 8.7/pH 7.0 ratio observed, down-regulated about 20-fold (Table 3). Some of the *che* and *mot* genes showed a relatively

TABLE 1. Clusters of pH-dependent genes

Cluster	Genes								
Acid High	<i>aceK</i>	<i>dapB</i>	<i>gatA</i>	<i>iap</i>	<i>murE</i>	<i>sirA (yhhP)</i>	<i>ybjN</i>	<i>ygdQ</i>	
	<i>acnA</i>	<i>dcyD (yedO)</i>	<i>gatB</i>	<i>icdA</i>	<i>murF</i>	<i>sprT</i>	<i>ycdB</i>	<i>yggA</i>	
	<i>acpD</i>	<i>dhaH^e (ycgC)</i>	<i>gatC</i>	<i>idi</i>	<i>nfnB</i>	<i>sthA (udhA)</i>	<i>ycdN</i>	<i>yggJ</i>	
	<i>add</i>	<i>dhaK^e (ycgT)</i>	<i>gatD</i>	<i>ispF (ygbB)</i>	<i>nfsA (mdaA)</i>	<i>sucA</i>	<i>ycdO^a</i>	<i>ygiW</i>	
	<i>ahpF</i>	<i>dhaL^e (ycgS)</i>	<i>gatY^{na,c}</i>	<i>kgtP</i>	<i>nuoC</i>	<i>sucB^a</i>	<i>ycfP</i>	<i>yhcN</i>	
	<i>alaT</i>	<i>edd</i>	<i>gatZ</i>	<i>lipA</i>	<i>nuoN</i>	<i>sucC^a</i>	<i>ycfR</i>	<i>yheN</i>	
	<i>alaU</i>	<i>eno</i>	<i>glcC</i>	<i>lldD</i>	<i>pdhR</i>	<i>sucD</i>	<i>ydgD</i>	<i>yheO</i>	
	<i>apbA</i>	<i>fdoG</i>	<i>glfA</i>	<i>lpxD</i>	<i>pdxY</i>	<i>tas</i>	<i>ydhM</i>	<i>yieE</i>	
	<i>aroA</i>	<i>fumA</i>	<i>glfB</i>	<i>lysC</i>	<i>proP</i>	<i>tehA</i>	<i>ydiH</i>	<i>yieF</i>	
	<i>aroH</i>	<i>fumC</i>	<i>glfD</i>	<i>lysU^a</i>	<i>rbsA</i>	<i>thrC</i>	<i>yeaS</i>	<i>yigI</i>	
	<i>aroP</i>	<i>fumI</i>	<i>gpmA</i>	<i>map</i>	<i>rbsC</i>	<i>trpB</i>	<i>yecD</i>	<i>yiiS^d</i>	
	<i>b0725</i>	<i>flgH</i>	<i>grxA</i>	<i>marA^a</i>	<i>rcsA</i>	<i>uspD (yiiT)</i>	<i>yecS</i>	<i>yjeM</i>	
	<i>b1364</i>	<i>flhA</i>	<i>gshA</i>	<i>marB</i>	<i>rimI</i>	<i>xseA</i>	<i>yejG</i>	<i>yjgK</i>	
	<i>bcsE (yjhS)</i>	<i>flhE</i>	<i>gshB</i>	<i>marR</i>	<i>rpoE</i>	<i>yabN</i>	<i>yfcA</i>	<i>yjiU</i>	
	<i>cadA^{a,b}</i>	<i>fliP</i>	<i>hdeA^{c,d}</i>	<i>mdaB</i>	<i>rseA</i>	<i>yagU</i>	<i>yfcD</i>	<i>ykgA</i>	
	<i>cfa^c</i>	<i>fliQ</i>	<i>hdeB^{c,d}</i>	<i>mdtG (yceE)</i>	<i>rseB</i>	<i>ybaK</i>	<i>yfcE</i>	<i>ylaC</i>	
	<i>cyaA</i>	<i>fliR</i>	<i>hemB</i>	<i>menF</i>	<i>sdhA</i>	<i>ybfd</i>	<i>yfid^{a,c}</i>	<i>yncD</i>	
	<i>cyoB</i>	<i>fliY^c</i>	<i>hsdR</i>	<i>metN (abc)</i>	<i>sdhB</i>	<i>ybgF</i>	<i>yjfF</i>	<i>yodC</i>	
	<i>dada</i>	<i>fumA</i>	<i>hsdS</i>	<i>mltC</i>	<i>serV</i>	<i>ybiC</i>	<i>yjfG</i>	<i>yohN</i>	
	<i>dadX</i>	<i>gapC</i>	<i>hslJ</i>	<i>mgo (yojH)</i>	<i>sfcA</i>	<i>ybjC</i>	<i>ygbE</i>	<i>yraQ</i>	
	Acid Low	<i>acrR</i>	<i>dcuS (yjdH)</i>	<i>gcvT</i>	<i>maa (ylaD)</i>	<i>ppiA</i>	<i>tatD (yigW)</i>	<i>yccA</i>	<i>yhaL</i>
		<i>abx^c (ygiT)</i>	<i>dksA^{c,e}</i>	<i>glgS</i>	<i>manX</i>	<i>pppA</i>	<i>tgt</i>	<i>yccK</i>	<i>yhiI</i>
		<i>atpH</i>	<i>dniR</i>	<i>glpA</i>	<i>mhpC</i>	<i>proV</i>	<i>tnaA^e</i>	<i>ycfS</i>	<i>yjgF^c</i>
		<i>b3913</i>	<i>dppC</i>	<i>hflB</i>	<i>miaA</i>	<i>purD</i>	<i>tnaB</i>	<i>ydHf</i>	<i>yjiA</i>
		<i>bax</i>	<i>dusA (yjbN)</i>	<i>hflC</i>	<i>mutL</i>	<i>purL</i>	<i>tnaC</i>	<i>yfiA</i>	<i>yjiX</i>
		<i>bioB</i>	<i>eco</i>	<i>hflK</i>	<i>nadA</i>	<i>purN</i>	<i>treB</i>	<i>ygaH</i>	<i>yjiY</i>
		<i>borD (ybcU)</i>	<i>fahD</i>	<i>hflX</i>	<i>nmpC</i>	<i>putA</i>	<i>treC</i>	<i>ygaU</i>	<i>ykfF</i>
<i>btuB</i>		<i>fhuD</i>	<i>hfpX</i>	<i>nrdD</i>	<i>recR</i>	<i>trs5</i>	<i>ygaZ</i>	<i>ylaB</i>	
<i>cirA</i>		<i>flpP</i>	<i>ilvB</i>	<i>nrdG</i>	<i>rpoH</i>	<i>ubiB (yigr)</i>	<i>ygdR</i>	<i>yibF</i>	
<i>cpxA</i>		<i>frdA</i>	<i>ilvG</i>	<i>nudH (ygdP)</i>	<i>rzpD (ybcT)</i>	<i>yagE</i>	<i>yggH</i>	<i>yncE</i>	
<i>cpxP^f</i>		<i>frdC</i>	<i>ilvN</i>	<i>ompF^e</i>	<i>sdaB</i>	<i>yahA</i>	<i>yghG</i>	<i>yqjA</i>	
<i>cpxR</i>		<i>fubB (yecI)</i>	<i>ivbL</i>	<i>phnA (yjdM)</i>	<i>sdaC</i>	<i>yajC</i>	<i>yghJ</i>	<i>yqjC</i>	
<i>dadX</i>		<i>fucI</i>	<i>katG</i>	<i>pitA</i>	<i>secD</i>	<i>ybaL</i>	<i>yghJ</i>	<i>yqjD</i>	
<i>dcuR (yjdG)</i>		<i>gcvH</i>	<i>kdtA</i>	<i>pntA</i>	<i>ssb^e</i>	<i>ybfA</i>	<i>ygiB</i>	<i>ytfJ</i>	
Base High		<i>art^c</i>	<i>b1171</i>	<i>dedA</i>	<i>glpX</i>	<i>osmB</i>	<i>recA</i>	<i>uvrY</i>	<i>yieG</i>
		<i>artM</i>	<i>b1172</i>	<i>dinI</i>	<i>glyA</i>	<i>potD</i>	<i>rnk</i>	<i>yceI^e</i>	<i>yihA</i>
		<i>asnA</i>	<i>b3837</i>	<i>dinJ</i>	<i>gpmB</i>	<i>prsA</i>	<i>rpiA^e</i>	<i>yciB</i>	<i>yijD</i>
	<i>aspA</i>	<i>carA</i>	<i>dsbA^e</i>	<i>himD</i>	<i>purA</i>	<i>sdaA^e</i>	<i>yciC</i>	<i>yjfO</i>	
	<i>atpA</i>	<i>coaA</i>	<i>emrR</i>	<i>hisC^c</i>	<i>purC</i>	<i>serU</i>	<i>yciI</i>	<i>yjgD</i>	
	<i>atpB</i>	<i>codB</i>	<i>eptB (yjhW)</i>	<i>hisF</i>	<i>purH</i>	<i>slt</i>	<i>ydcG</i>	<i>ynfD</i>	
	<i>atpC</i>	<i>cspD</i>	<i>fadL</i>	<i>hisH</i>	<i>purK</i>	<i>speA</i>	<i>ydeH</i>	<i>yqaE</i>	
	<i>atpD</i>	<i>cvpA</i>	<i>folD</i>	<i>hisI</i>	<i>purM</i>	<i>speD</i>	<i>yebE</i>	<i>yqgB</i>	
	<i>atpE</i>	<i>cydA</i>	<i>fucR</i>	<i>hisJ</i>	<i>pyrB</i>	<i>spy</i>	<i>yeel</i>	<i>yqjB</i>	
	<i>atpF</i>	<i>cydB</i>	<i>glmU</i>	<i>ispE (ychB)</i>	<i>pyrC</i>	<i>tatA</i>	<i>yehU</i>		
	<i>atpG</i>	<i>cydC</i>	<i>glpB</i>	<i>malT</i>	<i>pyrL</i>	<i>tatB</i>	<i>yfiQ</i>		
	<i>atpI</i>	<i>cydD</i>	<i>glpC</i>	<i>mdbO</i>	<i>rdoA (yihE)</i>	<i>ubiE (menG)</i>	<i>ygiC</i>		
	Base Low	<i>adhE^c</i>	<i>dnaI</i>	<i>flgM</i>	<i>fliS</i>	<i>ibpB^d</i>	<i>speG</i>	<i>ybcZ</i>	<i>yhhQ</i>
<i>aer</i>		<i>dnaK (ycgT)</i>	<i>flgN</i>	<i>fliT</i>	<i>lipB</i>	<i>srlA</i>	<i>ybgL</i>	<i>yhiN^d</i>	
<i>cca</i>		<i>dsbC</i>	<i>fliA</i>	<i>fliZ</i>	<i>lon</i>	<i>srlB</i>	<i>ybiX</i>	<i>yhjH</i>	
<i>cdd</i>		<i>fdoH</i>	<i>fliC</i>	<i>galM</i>	<i>modB</i>	<i>srlD</i>	<i>ybjX</i>	<i>yi8I</i>	
<i>cheA</i>		<i>flaG</i>	<i>fliD</i>	<i>gapA^e</i>	<i>motA</i>	<i>srlE</i>	<i>ycgR</i>	<i>yiaD</i>	
<i>cheB</i>		<i>flaX</i>	<i>fliE</i>	<i>gdhA</i>	<i>motB</i>	<i>srlR</i>	<i>ycjX</i>	<i>yjdA</i>	
<i>cheR</i>		<i>flgB</i>	<i>fliF</i>	<i>gntX (yhgH)</i>	<i>nhaB</i>	<i>tap</i>	<i>yeaD</i>	<i>ykfB</i>	
<i>cheW</i>		<i>flgC</i>	<i>fliG</i>	<i>groL (mopA)</i>	<i>nlpA</i>	<i>tar</i>	<i>yedW</i>	<i>ymdA</i>	
<i>cheY</i>		<i>flgD</i>	<i>fliH</i>	<i>groS (mopB)</i>	<i>nupG</i>	<i>tsr</i>	<i>yecR</i>	<i>yneE</i>	
<i>cheZ</i>		<i>flgE</i>	<i>fliI</i>	<i>grpE</i>	<i>ompT</i>	<i>tsx</i>	<i>yedM</i>	<i>ynfB</i>	
<i>clpB</i>		<i>flgF</i>	<i>fliJ</i>	<i>hlpA</i>	<i>pdxK^d</i>	<i>udp</i>	<i>yeeR</i>	<i>zntR (yhdM)</i>	
<i>clpX</i>		<i>flgG</i>	<i>fliK</i>	<i>hslO (yrfI)</i>	<i>prfC</i>	<i>yafE</i>	<i>yfbB</i>		
<i>deoA</i>		<i>flgI</i>	<i>fliL</i>	<i>hslU</i>	<i>rbsB</i>	<i>yafY</i>	<i>ygaZ</i>		
<i>deoB</i>		<i>flgJ</i>	<i>fliM</i>	<i>hslV</i>	<i>rbsD</i>	<i>ybbN</i>	<i>ygbF</i>		
<i>deoC</i>		<i>flgK</i>	<i>fliN</i>	<i>hspG</i>	<i>rbsK</i>	<i>ybeX</i>	<i>ygiS</i>		
<i>deoD</i>		<i>flgL</i>	<i>fliO</i>	<i>htrG (ygiM)</i>	<i>sfa (ymcE)</i>	<i>ybeY</i>	<i>yheL</i>		

Continued on following page

TABLE 1—Continued

Cluster	Genes								
Neutral High	<i>allA</i> (<i>ybbT</i>)	<i>frwB</i>	<i>idnT</i>	<i>menA</i>	<i>pmrD</i>	<i>uxaC</i>	<i>yhdT</i>	<i>yjhD</i>	
	<i>cdh</i>	<i>ftsA</i>	<i>lamB^e</i>	<i>mltB</i>	<i>psiF</i>	<i>yagF</i>	<i>yhfS</i>	<i>ykgF</i>	
	<i>dnaA</i>	<i>ftsQ</i>	<i>lnt</i>	<i>modC</i>	<i>pstA</i>	<i>ybaV</i>	<i>yhfU</i>	<i>ymdB</i>	
	<i>dnaN</i>	<i>fucK</i>	<i>lpxB</i>	<i>murG</i>	<i>pstB</i>	<i>ybcS</i>	<i>yhgE</i>	<i>yoaE</i>	
	<i>entE</i>	<i>galK</i>	<i>malE^{c,e}</i>	<i>napC</i>	<i>pstC</i>	<i>ybjG</i>	<i>yhiQ</i>	<i>yohK</i>	
	<i>fecA</i>	<i>gntT</i>	<i>malF</i>	<i>narY</i>	<i>pstS</i>	<i>ycdZ</i>	<i>yicG</i>	<i>yqfA</i>	
	<i>fecB</i>	<i>gpmM</i> (<i>yibO</i>)	<i>malG</i>	<i>nrdF</i>	<i>rne</i>	<i>ycjF</i>	<i>yihF</i>	<i>yqjH</i>	
	<i>fecC</i>	<i>gpsA</i>	<i>malK</i>	<i>pflC</i>	<i>tdcB</i>	<i>ycjO</i>	<i>yjaB</i>	<i>yrfF</i>	
	<i>fecD</i>	<i>hrpB</i>	<i>malM</i>	<i>pheA</i>	<i>tdcC</i>	<i>yddB</i>	<i>yjeJ</i>	<i>yrfG</i>	
	<i>fecE</i>	<i>hslR</i> (<i>yrfH</i>)	<i>malP</i>	<i>phoB</i>	<i>trkA</i>	<i>ydhF</i>	<i>yjflL</i>		
	<i>fepA</i>	<i>hupA</i>	<i>malQ</i>	<i>phoU</i>	<i>ulaE</i> (<i>sgaU</i>)	<i>yghD</i>	<i>yjgM</i>		
	<i>fhuC</i>	<i>hybA</i>	<i>mdlB</i>	<i>pioO</i> (<i>pioO</i>)	<i>ulaR</i> (<i>yjfQ</i>)	<i>yhaM</i>	<i>yjgW</i>		
	Neutral Low	<i>aceE</i>	<i>emrA</i>	<i>hpt</i>	<i>nadE</i>	<i>pflB^c</i>	<i>rho</i> (<i>sun</i>)	<i>ubiB</i>	<i>yezZ</i>
		<i>aceF^a</i>	<i>fabI</i>	<i>iadA</i>	<i>ndh</i>	<i>pflA</i>	<i>ribA</i>	<i>ubiH</i>	<i>yehS</i>
		<i>acnB</i>	<i>fadR</i>	<i>insA1</i>	<i>ndk</i>	<i>pflB^c</i>	<i>rmf</i>	<i>xerC</i>	<i>yehT</i>
		<i>adk</i>	<i>fdx</i>	<i>insA2</i>	<i>nemA</i>	<i>pgi</i>	<i>rnb</i>	<i>yadG</i>	<i>yeiG</i>
		<i>ahpC</i>	<i>fnr</i>	<i>kdsA</i>	<i>nuoG</i>	<i>pheM</i>	<i>rnt</i>	<i>yadH</i>	<i>yfaE</i>
<i>aldA</i>		<i>folE</i>	<i>kdsB</i>	<i>nuoH</i>	<i>pncB</i>	<i>rsd</i> (<i>yjaE</i>)	<i>yaiA</i>	<i>yfbQ</i>	
<i>apaG</i>		<i>fpr</i>	<i>lgt</i>	<i>nuoI</i>	<i>pps</i> (<i>ppsA</i>)	<i>rsuA</i>	<i>ybgC</i>	<i>yfcM</i>	
<i>argS</i>		<i>galF</i>	<i>lldP</i>	<i>nuoJ</i>	<i>ppx</i>	<i>sdhC</i>	<i>ybhB</i>	<i>yfcZ</i>	
<i>aspC</i>		<i>gapC</i>	<i>lpdA^d</i>	<i>nuoK</i>	<i>proC</i>	<i>sdhD</i>	<i>ycaR</i>	<i>yfdG</i>	
<i>avtA</i>		<i>ghrA</i> (<i>ycdW</i>)	<i>luxS^c</i> (<i>ygaG</i>)	<i>nuoL</i>	<i>pta^{c,e}</i>	<i>serC</i>	<i>ycbK</i>	<i>yfdI</i>	
<i>b0100</i>		<i>glf</i>	<i>lysP</i>	<i>ompR</i>	<i>ptsG</i>	<i>sfsB</i> (<i>nlp</i>)	<i>ycbL</i>	<i>yfhB</i>	
<i>can</i> (<i>yadF</i>)		<i>gloA</i>	<i>mdtJ</i>	<i>ompX^{a,e}</i>	<i>ptsH</i>	<i>sodB^d</i>	<i>ycdX</i>	<i>yfjW</i>	
<i>cll</i> (<i>wzzB</i>)		<i>glxX</i>	<i>mdlL</i>	<i>oppA^{c,e}</i>	<i>ptsI</i>	<i>sppA</i>	<i>ycdY</i>	<i>ygdl</i>	
<i>cyoA</i>		<i>gmhA</i>	<i>menB</i>	<i>oppB</i>	<i>ptsO</i>	<i>sra</i> (<i>rpsV</i>)	<i>yehH</i>	<i>yhbJ</i>	
<i>cyoC</i>		<i>gnd</i>	<i>mepA</i>	<i>oppC</i>	<i>purU</i>	<i>surA</i>	<i>yehJ</i>	<i>yheM</i>	
<i>cyoD</i>		<i>gppA</i>	<i>metG</i>	<i>oppD</i>	<i>pyrG</i>	<i>tatE</i> (<i>ybeC</i>)	<i>ydfG</i>	<i>yhjR</i>	
<i>cyoE</i>		<i>spt</i>	<i>metK</i>	<i>oppF</i>	<i>rbsB</i>	<i>tehB</i>	<i>ydiH</i>	<i>yieP</i>	
<i>cysK^c</i>		<i>grxB^c</i>	<i>miaB</i> (<i>yleA</i>)	<i>pal</i>	<i>relA</i>	<i>tolB</i>	<i>ydjN</i>	<i>yjbQ</i>	
<i>cysZ</i>		<i>guaA</i>	<i>mipA</i> (<i>yeaF</i>)	<i>pdxH</i>	<i>rfbC</i>	<i>tolQ</i>	<i>yeaC</i>	<i>yjeQ</i>	
<i>dld</i>		<i>guaB^d</i>	<i>mreB</i>	<i>pepB</i>	<i>rfbD</i>	<i>tpiA</i>	<i>yeaK</i>	<i>yjeS</i>	
<i>dps^c</i>		<i>hemM</i>	<i>mrp</i>	<i>pepN</i>	<i>rfe</i>	<i>tpx^{d,e}</i>	<i>yeaQ^e</i>	<i>yjgP</i>	
<i>dsbB</i>		<i>hha</i>	<i>mtn</i> (<i>pfs</i>)	<i>pepP</i>	<i>rhlB</i>	<i>typA</i> (<i>yihK</i>)	<i>yeaN</i>		

^a Acid induced (68, 73, 82).

^b Data from five arrays from pH 5.0; no significant expression at pH 7.0 and 8.7.

^c Acetate induced (3, 35).

^d Extreme acid resistance (44, 78, 79).

^e Base induced (68, 73, 82).

^f Base induced (16).

small degree of repression in acid compared to that at pH 7.0 but overall were repressed at high pH.

The major regulator operon *flhCD*, however, showed no effect of pH. Thus, either the *flhCD* probes failed to show up in our arrays or pH may affect expression posttranscriptionally.

Motility assays. The effect of pH on motility was tested by spotting motile cultures of *E. coli* K-12 RP437 and *S. enterica* serovar Typhimurium SJW1103 on motility agar buffered at a range of pH values (Fig. 4). Both species showed a steady decline of motility as pH increased. The decline was particularly steep between pH 7.5 and 8.7.

Catabolism and proton transport. Several enzymes for catabolism of sugars and amino acids show a pH dependence that minimizes acid production at low external pH or maximizes acids at high pH (68, 73). Our microarrays revealed many new components, showing the broad scope of pH regulation of catabolism (Table 4).

Many operons encoding processes of glycolysis and the TCA cycle, such as *aceEF* (pyruvate dehydrogenase), *dhaKL* (dihydroxyacetone kinase), *pta* (phosphotransacetylase), and *pts* (glucose phosphotransferase), showed elevated expression in acid. Others, however, were elevated at high pH. Operons elevated at high pH tended to be those induced by anaerobi-

osis, such as *glpABC* (anaerobic glycerol-3-phosphate dehydrogenase), *pflBA* (anaerobic pyruvate formate lyase), and *dcu* (anaerobic fumarate respiration). The *mal* system, however, is strongly repressed by acid (13, 31) and showed up as such in our arrays.

Membrane-bound systems for proton and electron transport were regulated by acid or base along lines largely consistent with their relative degree of export or import of H⁺. An example is the *atp* operon encoding F₁F_o ATP synthase (32), which imports H⁺ during oxidative respiration. Most of the *atp* genes were strongly upregulated at high pH, whereas *ndh* and *nuo* (the NADH dehydrogenases I and II), which export H⁺, were down-regulated. The *sdh* gene (succinate dehydrogenase), which contributes electrons for proton export, is also down-regulated at high pH. On the other hand, cytochrome *d* oxidase (*cyd*) is expressed in preference to cytochrome *o* oxidase (*cyo*) at high pH, presumably because it exports half as many H⁺ per electron (14).

Enzymes for degradation of amino acids showed pH regulation as expected, with high pH favoring deaminase operons such as *tna* (tryptophan deaminase), *sda* (serine deaminase), and *tdcB* (threonine dehydratase). Acid induced only one of the decarboxylase operons, *cad* (lysine decarboxylase). Several

TABLE 2. Strongest pH-dependent expression ratios (fourfold or higher)

pH dependence and pH ratio	Gene	Log ₂ ratio	pH dependence and pH ratio	Gene	Log ₂ ratio
Acid induced 5.0/7.0	<i>yagU</i>	3.220	Base induced 8.7/7.0	<i>yifO</i>	2.769
	<i>yhcN</i>	3.064		<i>ymgD</i>	2.221
	<i>sdhC</i>	2.728	8.7/5.0	<i>tnaC</i>	5.517
	<i>lysP</i>	2.662		<i>cpxP</i>	4.234
	<i>sdhD</i>	2.349		<i>tnaA</i>	4.028
	<i>cfa</i>	2.075		<i>nmpC</i>	3.961
	<i>nemaA</i>	2.060		<i>treB</i>	3.895
5.0/8.7	<i>yhcN</i>	4.199		<i>yjiY</i>	3.665
	<i>yagU</i>	3.962		<i>treC</i>	3.233
	<i>flhC</i>	3.396	<i>b3913</i>	3.176	
	<i>fimA</i>	2.579	<i>yifO</i>	3.095	
	<i>cfa</i>	2.555	<i>borD</i>	3.088	
	<i>gltB</i>	2.271	<i>tnaB</i>	2.993	
	<i>ydiY</i>	2.193	<i>ycfS</i>	2.820	
	<i>yedN</i>	2.147	<i>yghJ</i>	2.762	
	<i>yncD</i>	2.117	<i>ymgD</i>	2.433	
	<i>mgo</i>	2.075	<i>yccA</i>	2.378	
	<i>dhaH</i>	2.074	<i>yfiA</i>	2.364	
	<i>cheA</i>	2.074	<i>yebE</i>	2.343	
	<i>motB</i>	1.997	<i>yjiX</i>	2.292	
	7.0/8.7	<i>flhC</i>	4.561	<i>nrdD</i>	2.182
<i>malM</i>		3.780	<i>dniR</i>	2.081	
<i>malK</i>		3.748	<i>alx</i>	2.009	
<i>lamB</i>		3.735	<i>mutL</i>	2.000	
<i>malP</i>		3.373	7.0/5.0	<i>tnaC</i>	5.026
<i>motB</i>		3.238		<i>lamB</i>	4.881
<i>cheA</i>		3.199		<i>malK</i>	4.790
<i>cheZ</i>		3.199		<i>malM</i>	4.643
<i>flxA</i>		3.007		<i>yjiY</i>	4.359
<i>malE</i>		2.933		<i>nmpC</i>	4.012
<i>malQ</i>		2.883		<i>malP</i>	4.000
<i>cheW</i>		2.785		<i>tnaA</i>	3.805
<i>ibpB</i>		2.618		<i>malE</i>	3.425
<i>yhjH</i>		2.522		<i>borD</i>	3.378
<i>htpG</i>		2.439		<i>malQ</i>	3.322
<i>deoC</i>		2.267	<i>cpxP</i>	3.232	
<i>pstS</i>		2.262	<i>treB</i>	3.156	
<i>dnaK</i>		2.249	<i>yghJ</i>	3.081	
<i>tar</i>		2.213	<i>yjiX</i>	2.959	
<i>yjdA</i>		2.208	<i>fecB</i>	2.842	
<i>dnaJ</i>		2.155	<i>ompF</i>	2.834	
<i>yrfG</i>		2.055	<i>treC</i>	2.613	
<i>yheL</i>		2.031	<i>fecA</i>	2.608	
<i>deoA</i>	2.003	<i>pstS</i>	2.479		
		<i>b3913</i>	2.177		
		<i>fecE</i>	2.098		

decarboxylases are known to be induced by acid, but their induction is repressed by oxygen (4, 30), which may explain their absence in our highly aerobic cultures.

Oxidative stress and salicylate stress. Several acid stress genes are known to overlap with oxidative stress, for example, the alkyl hydroperoxide reductase *ahpC* (9, 84), and certain permeant acids such as salicylate are considered oxidative stress agents (54). We surveyed our pH-regulated genes for overlap with response to H₂O₂, paraquat, and salicylate, as reported in references 54 and 84 (Table 5).

Of the 73 pH-dependent genes known to be induced by H₂O₂, paraquat, or salicylate, virtually all were induced by acid or repressed by base. This finding confirms our hypothesis of a strong connection between acid stress and oxidative stress. It may be that low pH amplifies the toxicity of oxygen radicals.

Genes repressed by paraquat or salicylate were repressed in acid or induced at high pH, such as the base-inducible membrane protein gene *alx*, the histidine cyclase gene *hisF*, and outer membrane protein gene *ompF*. An exception to these generalizations was the maltose regulon (*lamB*, *malE*, and *malK*), which was repressed by acid but induced by paraquat.

Envelope and periplasmic stress. A large part of *E. coli* function takes place in the outer membrane and envelope (48) and the periplasm (49), compartments essentially exposed to “extracellular” pH. Thus, it is not surprising that several envelope and periplasmic components show pH-dependent expression (16, 23, 73, 82). Our microarrays revealed an even greater number of such responses (Table 6). Both acid and base induction were observed. Acid-induced periplasmic proteins in-

TABLE 3. Flagellar and chemotaxis genes

Gene	Function	Log ₂ pH ratio ^a			Class ^b
		5/7	8.7/7	8.7/5	
<i>cheA</i>	Chemotaxis sensor kinase	-1.125	-3.199	-2.0474	BL
<i>cheB</i>	Protein methylesterase	-1.050	-1.578	-0.528	BL
<i>cheR</i>	Chemotaxis MCP ^c methyltransferase	-0.564	-1.013	-0.448	BL
<i>cheW</i>	Chemotaxis signal transducer	-1.336	-2.785	-1.449	BL
<i>cheY</i>	Response regulator for chemotactic signal	-1.089	-1.310	-0.221	BL
<i>cheZ</i>	CheY-P phosphatase	-1.505	-3.199	-1.694	BL
<i>flgA</i>	Flagellar synthesis	-0.261	-1.056	-0.795	BL
<i>flgB</i>	Basal body rod subunit	-0.192	-1.120	-0.928	BL
<i>flgC</i>	Basal body rod subunit	-0.257	-1.241	-0.984	BL
<i>flgD</i>	Basal body rod modification	0.133	-1.241	-1.107	BL
<i>flgE</i>	Hook subunit	0.272	-0.856	-1.128	BL
<i>flgF</i>	Basal body rod subunit	0.109	-1.239	-1.348	BL
<i>flgG</i>	Basal body rod major subunit	0.220	-1.116	-1.335	BL
<i>flgI</i>	Basal body P-ring	-0.011	-1.218	-1.207	BL
<i>flgJ</i>	Flagellum-specific muramidase	0.030	-0.857	-0.886	BL
<i>flgK</i>	Flagellar synthesis	-0.211	-1.875	-1.664	BL
<i>flgL</i>	Flagellar synthesis	0.111	-1.165	-1.276	BL
<i>flgM</i>	Anti-sigma 28 (FliA); regulates FlhD	-0.292	-1.424	-1.132	BL
<i>flgN</i>	Flagellar synthesis	-0.295	-1.567	-1.272	BL
<i>flhA</i>	Flagellar export pore protein	0.333	-0.528	-0.861	AH
<i>flhE</i>	Function unknown	0.492	-0.454	-0.946	AH
<i>fliA</i>	Sigma 28; regulates class III flagellar genes	-0.150	-1.127	-0.976	BL
<i>fliC</i>	Flagellin subunit, H-antigen	-1.165	-4.561	-3.396	BL
<i>fliD</i>	Hook-associated protein	-0.311	-1.953	-1.641	BL
<i>fliE</i>	Flagellar synthesis; basal body component	-0.478	-1.861	-1.203	BL
<i>fliF</i>	Flagellar basal body M-ring	-0.159	-1.216	-1.057	BL
<i>fliG</i>	Motor switching and energizing	0.182	-1.313	-1.495	BL
<i>fliH</i>	Negative regulator of FliI; flagellar assembly and export	0.389	-1.245	-1.634	BL
<i>fliI</i>	Membrane ATPase, flagellar, axial subunit export	0.270	-1.238	1.508	BL
<i>fliJ</i>	Flagellar biosynthesis	-0.068	-1.404	-1.336	BL
<i>fliK</i>	Hook filament junction	0.278	-1.186	-1.464	BL
<i>fliL</i>	Rotational direction of flagella	-0.102	-1.083	-0.981	BL
<i>fliM</i>	Flagellar synthesis, motor switching and energizing	0.029	-1.053	-1.082	BL
<i>fliN</i>	Flagellar switch	0.273	-0.875	-1.148	BL
<i>fliO</i>	Flagellar synthesis	0.191	-0.921	-1.112	BL
<i>fliP</i>	Flagellar synthesis	0.419	-0.669	-1.087	AH
<i>fliQ</i>	Flagellar synthesis	0.463	-0.980	-1.444	AH
<i>fliR</i>	Flagellar synthesis	0.924	-0.899	-1.822	AH
<i>fliS</i>	Cytosolic chaperone inhibits premature FliC assembly	-0.174	-1.947	-1.772	BL
<i>fliT</i>	Flagellar synthesis	-0.323	-1.021	-0.698	BL
<i>fliY</i>	Cystine-binding protein, periplasmic; may regulate FliA (sigma 28)	0.233	-0.252	-0.484	AH
<i>fliZ</i>	Not required for motility; may regulate FliA (sigma 28)	0.144	-1.165	-1.309	BL
<i>motA</i>	Flagellar rotation	-0.682	-1.847	-1.166	BL
<i>motB</i>	Flagellar rotation	-1.241	-3.238	-1.997	BL
<i>tap</i>	Dipeptide chemoreceptor	-0.622	-1.089	-0.466	BL
<i>tar</i>	Aspartate, maltose chemoreceptor	-0.519	-2.213	-1.694	BL
<i>tsr</i>	Serine chemoreceptor	-1.061	-1.826	-0.765	BL
<i>ycgR</i>	Suppresses <i>hms</i> motility defect	-0.531	-0.783	-0.252	BL
<i>yhjH</i>	Suppresses <i>hms</i> motility defect	-0.972	-2.522	-1.550	BL

^a Boldface for ratios indicates significance ($\alpha = 0.001$).

^b BL, Base Low; AH, Acid High.

^c MCP, methyl-accepting chemotaxis protein.

cluded the well-known acid chaperone from *hdeAB* (23), as well as the newly observed TolA-binding protein (*ybgF*) and the lipoprotein from *pal*. High pH induced the ferric transporters from *fecAB* and *fluD*, possibly due to low iron solubility at high pH. At high pH, various transport proteins and redox modulators such as that from *dsbA* are known to be induced. In addition, several additional base-induced periplasmic and envelope proteins appeared, including the vitamin B₁₂ transporter from *btuB*, the outer membrane protein

from *nmpC*, and the peptidylprolyl-*cis-trans*-isomerase from *ppiA*.

Universal stress and heat shock. Various heat shock and universal stress proteins are inducible by the permeant acid benzoate, such as the products of *clpB*, *htpG*, *dnaK*, *groS*, and *uspA* (38). Some of these showed pH response in our microarrays (Table 7). The DNA damage response gene *uspD* was acid induced, as was *dps*, encoding the DNA-binding protein involved in stationary phase and acid resistance. Acid induced

rseAB, the antisigma regulators of the *rpoE* envelope heat stress system (1). High pH induced the *rpoH* heat shock sigma 32 gene (28) as well as heat shock proteasome genes *hslUV* and regulators *hslOR*.

DISCUSSION

Overall, our work revealed a large number of genes not previously known to be regulated by pH. Furthermore, many of these genes had no previously known function or response, such as *yhcN* and *yagU* (induced by acid) and *yifO* and *ymcG* (induced by base).

An important question is to assess the biological relevance of the expression ratios reported (36, 51). Most of the ratios we reported as significant (boldface in Tables 3 through 7) are greater than twofold ($\log_2 = 1$). In many cases, all or most members of an operon fell in the same cluster and show similar expression profiles; the flagellar regulon was particularly consistent (Table 3). The gene probes are synthesized on the array independently of their operon map; thus, parallel expression profiles within operons do not reflect array position. Note that even genes with significant expression ratios of less than 2 ($\log_2 = 1$) tend to group with their operons. In previous studies, comparison with quantitative reverse transcriptase real-time PCR shows that microarray ratios, while quantitatively consistent, generally underestimate the actual differences in mRNA levels between the biological systems compared (83).

Flagellar biosynthesis and motility. The effects of pH in flagellar biosynthesis and motility remain poorly understood. It has long been known that low external pH (thus, large Δ pH) contributes to the proton motive force that drives flagellar rotation (33). The cytoplasmic pH, however, must remain high; permeant acids such as acetate and benzoate, which depress internal pH and decrease proton motive force, are chemotactic repellents (67) and impair rotation of the flagellar motor (46). Low pH elicits negative chemotaxis (55, 67), whereas a pH increase up to 8.3 elicits a positive response (55).

In recent reports acid stress is associated with low motility (72), yet acetate has been reported to induce the flagellar regulon and enhance motility (53). We believe that the previous reports are limited in several ways. Reference 72 does not compare pH conditions directly but notes repression of flagellar genes in an *hms* mutant in which acid resistance is increased. The motility assay is not clearly described, and the acid dependence of *flhDC-cat* expression was observed on plasmids, not in the genome. Reference 53 reports induction of chromosomal *flhDC-lacZ* fusions by acetate. Those authors' assays of motility, however, show relatively small differences between pH conditions.

Our microarrays showed strong evidence for suppression of motility and chemotaxis at high pH. This evidence was supported by the decrease in motility at high pH, observed for both *E. coli* and *S. enterica* serovar Typhimurium, which swims twice as fast as *E. coli*. We also found weaker evidence for repression of *che* and *mot* genes at pH 5, but the flagellar synthesis genes were strongly induced at low pH. Overall, our data point to alkaline suppression of flagellar motility. Work in progress shows that, at high pH, the number of flagella per cell is decreased to one to three per cell (about 20% of normal) (*S. Aizawa and J. Slonczewski, unpublished data*).

No pH dependence was observed for the flagellar regulators

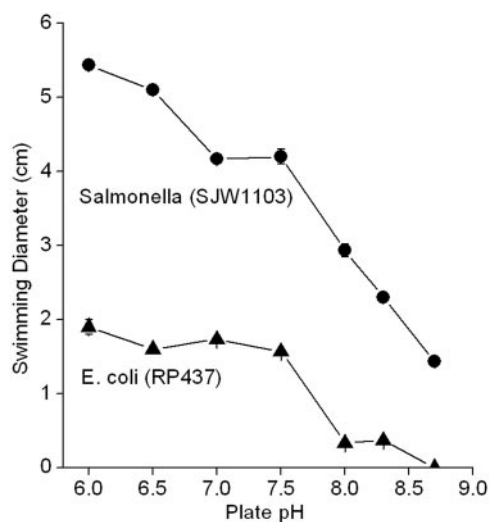


FIG. 4. Swimming distance as a function of pH. *E. coli* K-12 RP437 and *S. enterica* serovar Typhimurium SJW1103 were spotted on soft-agar plates as described under Materials and Methods. Error bars represent standard errors of the means ($n = 3$); in most cases their size was smaller than the symbol.

flhD and *flhC*. On the other hand, in a microarray study of anaerobic cultures, *flhD* and *flhC* are induced by acid (*E. Hayes and J. L. Slonczewski, unpublished data*). Acid induction of these regulators would be consistent with the report of their induction by acetate (53). We did see acid induction of two known activators of *flhDC*: adenylate cyclase *cyaA* (37) (Acid High) and *dnaK-dnaJ-grpE* (64) (Base Low). We saw no acid induction of other flagellar activators such as *crp* (37), nor did we see alkaline induction of the negative flagellar regulator *rcsCDB* (22).

An alternative model is that pH regulation of the flagellar regulon is mediated by proteolysis, as in the case of ClpXP proteolysis of FlhD and FlhC (76). We find that ClpX is down-regulated at high pH (Base Low cluster), but a different protease could be involved.

Catabolism. The picture of catabolism is more complicated, but in general our expression ratios confirm our present hypotheses of pH regulation while extending our knowledge to many more components. Systems that consume acids are enhanced at low pH. On the other hand, initial import and breakdown of some sugars, such as maltose, are favored at high pH, where they may quickly generate a large burst of fermentation acids.

With respect to proton export, *E. coli* appears to prefer components such as ATP synthase that import protons at high pH (counteracting the alkaline stress on cytoplasmic pH) and prefers to minimize proton export associated with the terminal oxidase *cyd* in preference to *cyo*. This observation is consistent with the previous report that *cyd* expression is higher at pH 7.5 than at pH 5.0 in an *fnr* mutant (14), although in those experiments *cyo* expression also increased with pH. It is likely that our broader range of pH classes (up to pH 8.7) provided a clearer picture of pH regulation of *cyo* and *cyd*.

Under amino acid catabolism, relatively few new components of pH response were observed. This makes sense, because most amino acid decarboxylases are repressed by oxygen

TABLE 4. Catabolism and respiration

Group	Gene	Function	Log ₂ pH ratio ^d			Class ^b
			5/7	8.7/7	8.7/5	
Sugar catabolism and TCA cycle	<i>aceE</i>	Pyruvate dehydrogenase	1.928	0.527	-1.401	NL
	<i>aceF</i>	Pyruvate dehydrogenase dihydrolipoamide acetyltransferase	1.802	0.402	1.401	NL
	<i>aceK</i>	Isocitrate dehydrogenase kinase/phosphatase	0.667	0.038	-0.629	AH
	<i>acnA</i>	Aconitase A, stationary phase induced	0.769	-0.143	-0.912	AH
	<i>acnB</i>	Aconitase B; 2-methylaconitate hydratase	1.036	-0.041	-1.077	NL
	<i>acrR</i>	Regulator for <i>acrA</i> and <i>acrB</i>	-0.873	0.164	1.037	AL
	<i>dcuR</i> (<i>yjdG</i>)	Fumarate respiration regulator (anaerobic)	-0.220	0.055	0.275	AL
	<i>dcuS</i> (<i>yjdH</i>)	Fumarate respiration regulator (anaerobic)	-0.324	0.065	0.390	AL
	<i>dhaK</i>	Dihydroxyacetone kinase, subunit I	1.041	-0.379	-1.420	AH
	<i>dhaL</i>	Dihydroxyacetone kinase, subunit II	0.742	-0.385	-1.127	AH
	<i>dld</i>	D-Lactate dehydrogenase	0.919	0.566	-0.353	NL
	<i>eno</i>	Enolase; RNA degradosome	0.299	0.052	-0.247	AH
	<i>fucI</i>	L-Fucose isomerase	-0.693	-0.173	0.520	AL
	<i>fucK</i>	L-Fuculose kinase	-1.259	-0.519	0.740	NH
	<i>fucR</i>	Positive regulator, <i>fuc</i> operon	-0.046	0.495	0.541	BH
	<i>galF</i>	Putative regulator of <i>galU</i>				NL
	<i>galK</i>	Galactokinase	-0.616	-0.562	0.053	BL
	<i>galM</i>	Galactose mutarotase; aldose-1-epimerase	-0.619	-0.647	-0.028	
	<i>gapA</i>	Glyceraldehyde 3-P dehydrogenase A	0.093	-0.752	-0.844	BL
	<i>gatA</i>	Galactitol-specific enzyme IIA of PTS ^d	0.929	-0.626	-1.555	AH
	<i>gatB</i>	Galactitol-specific enzyme IIB of PTS	0.632	-0.895	-1.528	AH
	<i>gatC</i>	Galactitol-specific enzyme IIC of PTS	0.973	-0.949	-1.922	AH
	<i>gatD</i>	Galactitol-1-phosphate dehydrogenase	1.037	-0.893	-1.930	AH
	<i>gatY</i>	D-Tagatose-1,6-bisphosphate aldolase, class II	0.846	-0.483	-1.329	AH
	<i>gatZ</i>	Enhances GatY activity	0.774	-0.605	-1.380	AH
	<i>glpA</i>	Glycerol-3-phosphate dehydrogenase large subunit (anaerobic)	-0.200	0.558	0.758	AL
	<i>glpB</i>	Glycerol-3-phosphate membrane anchor (anaerobic)	-0.124	0.437	0.562	BH
	<i>glpC</i>	Glycerol-3-phosphate dehydrogenase (anaerobic) small subunit	-0.169	0.469	0.638	BH
	<i>glpX</i>	Fructose 1,6-bisphosphatase	-0.162	0.375	0.537	BH
	<i>gltA</i>	Citrate synthase	0.288	-0.559	-0.846	AH
	<i>gnd</i>	Gluconate-6-phosphate dehydrogenase	0.895	0.351	-0.544	NL
	<i>gntT</i>	High-affinity gluconate transport	-0.613	-0.362	0.251	NH
	<i>gpsA</i>	Glycerol-3-phosphate dehydrogenase	-0.492	-0.534	-0.041	NH
	<i>gpmA</i>	Phosphoglycerate mutase I	0.303	-0.118	-0.421	AH
	<i>icdA</i>	Isocitrate dehydrogenase	1.211	0.061	-1.272	AH
	<i>lldD</i>	L-Lactate dehydrogenase	0.613	-0.554	-1.167	AH
	<i>lldP</i>	L-Lactate permease; glycolate uptake	1.527	0.219	-1.307	NL
	<i>lpdA</i>	Lipoamide dehydrogenase; E3 component of pyruvate and 2-oxoglutarate dehydrogenase complexes	1.507	0.281	-1.226	NL
	<i>malE</i>	Maltose-binding protein, periplasmic	-3.425	-2.933	0.491	NH
	<i>malF</i>	Maltose transport, inner membrane	-1.016	-0.935	0.082	NH
	<i>malG</i>	Maltose transport, inner membrane subunit	-1.577	-1.502	0.075	NH
	<i>malK</i>	Maltose transport, ATP-binding subunit	-4.790	-3.748	1.041	NH
	<i>malM</i>	Periplasmic protein, <i>mal</i> regulon	-4.643	-3.780	0.863	NH
	<i>malP</i>	Maltodextrin phosphorylase	-4.000	-3.373	0.627	NH
	<i>malQ</i>	Amylomaltase	-3.322	-2.883	0.439	NH
	<i>malT</i>	<i>mal</i> positive regulator	-0.103	0.617	0.720	BH
	<i>pdhR</i>	Pyruvate dehydrogenase operon repressor	0.755	-0.109	-0.863	AH
	<i>pfkB</i>	6-Phosphofructokinase-2	0.363	0.095	-0.268	NL
	<i>pflA</i>	Pyruvate formate lyase I activase	0.964	0.667	-0.297	NL
	<i>pflB</i>	Pyruvate formate lyase I (anaerobic)	0.812	0.578	-0.234	NL
	<i>pgi</i>	Glucose phosphate isomerase	0.654	0.177	-0.477	NL
	<i>pta</i>	Phosphotransacetylase	1.167	0.639	-0.528	NL
	<i>ptsG</i>	Glucose PTS enzyme IIBC	0.841	0.544	-0.287	NL
	<i>ptsH</i>	PTS system histidine phosphocarrier protein Hpr	0.369	0.564	0.195	NL
	<i>ptsI</i>	PTS system enzyme I	0.241	0.339	0.098	NL
	<i>ptsO</i>	NPr, N-regulated HPr-like protein	0.639	0.168	-0.470	NL
	<i>rpiA</i>	Ribose-5-phosphate isomerase A	0.180	0.364	0.184	BH
	<i>srlA</i>	Sorbitol-specific enzyme II of PTS	-1.156	-1.720	-0.564	BL
	<i>srlB</i>	Sorbitol-specific enzyme III of PTS	-0.908	-1.140	-0.232	BL
	<i>srlD</i>	Sorbitol-6-phosphate dehydrogenase	-0.660	-1.334	-0.674	BL
	<i>srlE</i>	srl operon protein	-1.013	-1.613	-0.599	BL
	<i>srlR</i>	srl regulator	-0.151	0.804	-0.654	BL
	<i>sucA</i>	2-Oxoglutarate dehydrogenase, E1 component	0.850	-0.851	-1.701	AH
	<i>sucB</i>	Dihydrolipoamide succinyltransferase component of 2-oxoglutarate dehydrogenase complex (E2)	0.566	-1.073	-1.638	AH
	<i>sucC</i>	Succinyl-CoA ^a synthase beta subunit	0.494	-1.019	-1.513	AH
<i>sucD</i>	Succinyl-CoA synthase alpha subunit	0.526	-1.225	-1.751	AH	
<i>tpiA</i>	Triosephosphate isomerase	0.772	0.245	-0.526	NL	

Continued on following page

Downloaded from http://jib.asm.org/ on September 27, 2015 by guest

TABLE 4—Continued

Group	Gene	Function	Log ₂ pH ratio ^a			Class ^b
			5/7	8.7/7	8.7/5	
Proton transport and electron transport chain	<i>treB</i>	Trehalose-specific PTS enzyme II	-3.156	0.739	3.895	AL
	<i>treC</i>	Trehalose-6-phosphate hydrolase	-2.613	0.620	3.233	AL
	<i>atpA</i>	ATP synthase subunit alpha, F ₁	0.111	0.460	0.349	BH
	<i>atpB</i>	ATP synthase subunit a, F ₀	0.125	0.484	0.609	BH
	<i>atpC</i>	ATP synthase subunit epsilon, F ₁	0.266	1.005	0.739	BH
	<i>atpD</i>	ATP synthase subunit beta, F ₁	0.542	1.172	0.630	BH
	<i>atpE</i>	ATP synthase subunit c, F ₀	0.182	0.503	0.321	BH
	<i>atpF</i>	ATP synthase subunit b, F ₀	-0.055	0.321	0.377	BH
	<i>atpG</i>	ATP synthase subunit gamma, F ₁	0.372	0.632	0.260	BH
	<i>atpH</i>	ATP synthase subunit delta, F ₁	-0.172	0.186	0.358	AL
	<i>atpI</i>	ATP synthase subunit, F ₁ F ₀ -type proton-ATPase	0.017	0.499	0.482	BH
	<i>cydA</i>	Cytochrome <i>d</i> (bd-I) terminal oxidase subunit I (microaerobic)	-0.268	0.872	1.140	BH
	<i>cydB</i>	Cytochrome <i>d</i> (bd-I) terminal oxidase subunit II (microaerobic)	-0.097	0.777	0.874	BH
	<i>cydC</i>	Cysteine exporter to periplasm required for Cyd assembly	0.135	0.490	0.355	BH
	<i>cydD</i>	Cysteine exporter to periplasm required for cytochrome assembly	0.232	0.574	0.342	BH
	<i>cyoA</i>	Cytochrome <i>o</i> oxidase subunit II	1.160	0.217	-0.943	NL
	<i>cyoB</i>	Cytochrome <i>o</i> oxidase subunit I	0.812	-0.204	-1.016	NL
	<i>cyoC</i>	Cytochrome <i>o</i> oxidase subunit III	1.026	0.019	-1.007	NL
	<i>cyoD</i>	Cytochrome <i>o</i> oxidase subunit IV	0.829	-0.192	-1.021	NL
	<i>cyoE</i>	Cytochrome <i>o</i> oxidase subunit protoheme IX farnesyltransferase	1.094	0.225	-0.869	NL
	<i>fdoG</i>	Formate dehydrogenase-O, major selenopeptide subunit	0.353	-0.138	-0.491	AH
	<i>fdoH</i>	Formate dehydrogenase-O Fe-S subunit	0.003	-0.414	-0.416	BL
	<i>frdA</i>	Fumarate reductase flavoprotein subunit	-0.305	0.046	0.351	AL
	<i>frdC</i>	Fumarate reductase membrane anchor polypeptide	-0.378	-0.008	0.370	AL
	<i>fumA</i>	Fumarase A	0.595	-0.647	-1.242	AH
	<i>nfsA (mdaA)</i>	Nitroreductase A	0.561	-0.419	-0.980	AH
	<i>mdaB</i>	Probable nitroreductase or quinone reductase	0.444	-0.223	-0.667	AH
	<i>napC</i>	Cytochrome electron source for NapAB, membrane bound	-0.797	-0.382	0.416	NH
	<i>ndh</i>	Respiratory NADH dehydrogenase II; NADH:ubiquinone oxidoreductase II	0.789	0.161	-0.628	NL
	<i>nuoC</i>	NADH:ubiquinone oxidoreductase subunit C	0.440	-0.084	-0.524	AH
	<i>nuoG</i>	NADH:ubiquinone oxidoreductase subunit G; NADH dehydrogenase I	0.525	0.103	-0.442	NL
	<i>nuoH</i>	NADH:ubiquinone oxidoreductase subunit H; NADH dehydrogenase I	0.564	0.185	-0.379	NL
	<i>nuoI</i>	NADH:ubiquinone oxidoreductase subunit I; NADH dehydrogenase I	0.792	0.310	-0.481	NL
	<i>nuoJ</i>	NADH:ubiquinone oxidoreductase subunit J; NADH dehydrogenase I	0.468	0.234	-0.234	NL
	<i>nuoK</i>	NADH:ubiquinone oxidoreductase subunit K; NADH dehydrogenase I	0.766	0.460	-0.306	NL
	<i>nuoL</i>	NADH:ubiquinone oxidoreductase subunit L; NADH dehydrogenase I	0.311	0.078	-0.233	NL
<i>nuoN</i>	NADH:ubiquinone oxidoreductase subunit N; NADH dehydrogenase I	0.126	-0.210	-0.336	AH	
<i>sdhA</i>	Succinate dehydrogenase flavoprotein subunit	1.438	-0.018	-1.458	AH	
<i>sdhB</i>	Succinate dehydrogenase iron-sulfur protein	1.833	0.329	-1.505	AH	
<i>sdhC</i>	Succinate dehydrogenase membrane anchor subunit, cytochrome <i>b</i> ₅₅₆	2.728	0.890	-1.838	NL	
<i>sdhD</i>	Succinate dehydrogenase hydrophobic subunit	2.349	0.595	-1.754	NL	
Amino acid catabolism and transport	<i>artI</i>	Arginine periplasmic binding protein	0.286	0.655	0.369	BH
	<i>artM</i>	Arginine periplasmic binding protein	-0.115	0.293	0.407	BH
	<i>cadA</i>	Lysine decarboxylase, degradative	1.024	0.137	-0.887	AH
	<i>cysK</i>	<i>O</i> -Acetylserine sulfhydrylase A (cysteine synthase)	1.204	1.351	0.147	NL
	<i>dadA</i>	D-Amino acid dehydrogenase	1.273	-0.229	-1.501	AH
	<i>dadX</i>	D-Amino acid dehydrogenase	0.683	-0.393	-1.076	AH
	<i>dppC</i>	Dipeptide permease system	-0.346	-0.032	0.315	AL
	<i>gdhA</i>	Glutamate dehydrogenase	-0.080	-0.496	-0.416	BL
	<i>hisC</i>	Histidinol-phosphate aminotransferase	0.253	0.984	0.731	BH
	<i>hisF</i>	Imidazole glycerol phosphate synthase (cyclase)	0.150	0.703	0.553	BH
	<i>hisH</i>	Amidotransferase of imidazole glycerol phosphate synthase	0.034	0.393	0.359	BH
	<i>hisI</i>	PR-ATP pyrophosphatase and PR-AMP cyclohydrolase	0.234	0.641	0.407	BH
	<i>hisJ</i>	Histidine-binding protein	-0.149	0.400	0.549	BH
	<i>lysC</i>	Aspartokinase III	1.464	-0.226	-1.690	AH
	<i>lysP</i>	Lysine permease	2.662	0.963	-1.698	NL
	<i>lysU</i>	Lysine-tRNA ligase	0.544	-0.077	-0.621	AH
	<i>potD</i>	Putrescine-ornithine transporter	0.053	0.509	0.456	BH
	<i>sdaA</i>	L-Serine deaminase, degradative	-0.409	0.638	1.048	BH
	<i>sdaB</i>	L-Serine deaminase	-1.111	-0.434	0.678	AL
	<i>sdaC</i>	H ⁺ /serine symporter; regulator of serine deaminase	-1.205	-0.410	0.794	AL
<i>tnaA</i>	Tryptophan deaminase, degradative; also deaminases serine and cysteine	-3.805	0.223	4.028	AL	
<i>tnaB</i>	Tryptophan transporter	-1.840	1.153	2.993	AL	
<i>tnaC</i>	<i>tnaA</i> leader peptide	-5.026	0.490	5.517	AL	
<i>tdcB</i>	Threonine dehydratase, degradative	-0.849	-0.296	0.553	NH	
<i>ydfG</i>	L-allo-Threonine, L-serine, D-serine dehydrogenase	0.407	0.287	-0.120	NL	

^a Values in boldface are significant ($\alpha = 0.001$).

^b NL, Neutral Low; AH, Acid High; AL, Acid Low; BH, Base High; BL, Base Low; NH, Neutral High.

^c CoA, coenzyme A.

^d PTS, phosphotransferase.

TABLE 5. pH-regulated oxidative stress response^a

Gene	Function	Log ₂ pH ratio ^b			PQ, Sal, or H ₂ O ₂	Class ^c
		5/7	8.7/7	8.7/5		
<i>acnA</i>	Aconitase A	0.769	-0.143	-0.912	Sal	AH
<i>adhE</i>	Acetaldehyde-coenzyme A dehydrogenase	0.043	-0.380	-0.422	Sal	BL
<i>ahpC</i>	Alkyl hydroperoxide reductase small subunit	1.003	0.436	-0.568	PQ, H ₂ O ₂	NL
<i>ahpF</i>	Alkyl hydroperoxide reductase large subunit	0.777	-0.222	-0.999	H ₂ O ₂	AH
<i>aldA</i>	Aldehyde dehydrogenase, NAD linked	0.773	0.764	-0.009	PQ	NL
<i>alx</i> (<i>vgjT</i>)	Membrane protein, alkali induced	-1.317	0.692	2.009	PQ-	AL
<i>artI</i>	Periplasmic arginine binding protein	0.286	0.655	0.369	PQ	BH
<i>aspA</i>	Aspartate ammonia-lyase (aspartase)	-0.592	0.770	1.362	PQ-	BH
<i>carA</i>	Carbamoylphosphate synthase small subunit	0.029	1.059	1.030	PQ-	BH
<i>cfa</i>	Cyclopropane fatty acid synthase	2.075	-0.480	-2.555	Sal	AH
<i>cyaA</i>	Adenylate cyclase	0.691	-0.156	-0.846	Sal	AH
<i>cyoD</i>	Cytochrome <i>o</i> oxidase subunit IV	0.829	-0.192	-1.021	PQ	NL
<i>cysK</i>	Cysteine synthase	1.204	1.351	0.147	PQ, Sal, H ₂ O ₂	NL
<i>dadX</i>	Alanine racemase	0.683	-0.393	-1.076	PQ	AH
<i>deoA</i>	Thymidine phosphorylase	-1.031	-2.003	-0.972	Sal	BL
<i>deoB</i>	Deoxyrioubourate, phosphopentomutase	-0.731	-1.502	-0.771	PQ, Sal	BL
<i>dhaH</i>	Dihydroxyacetone phosphoryl donor	1.547	-0.527	-2.074	Sal	AH
<i>dhaK</i>	Dihydroxyacetone kinase	1.041	-0.379	-1.420	Sal	AH
<i>dnaK</i>	HSP-70-type molecular chaperone	-0.894	-2.249	-1.356	Sal	BL
<i>dps</i>	Stress response DNA-binding protein	1.130	0.105	-1.025	PQ, Sal, H ₂ O ₂	NL
<i>fltS</i>	Flagellar synthesis; flagellar regulon member	-0.174	-1.947	-1.772	PQ-	BL
<i>fpr</i>	Ferredoxin NADP ⁺ reductase; anaerobic	0.565	0.275	-0.289	PQ, H ₂ O ₂	NL
<i>gapA</i>	GAPDH ^d A	0.093	-0.752	-0.844	Sal	BL
<i>gatA</i>	Galactitol-specific enzyme IIA of PTS ^e	0.929	-0.626	-1.555	PQ, Sal	AH
<i>gatB</i>	Galactitol-specific enzyme IIB of PTS	0.632	-0.895	-1.528	PQ, Sal	AH
<i>gatC</i>	Galactitol-specific enzyme IIC of PTS	0.973	-0.949	-1.922	Sal	AH
<i>gatD</i>	Galactitol-1-phosphate dehydrogenase	1.037	-0.893	-1.930	PQ, Sal	AH
<i>gatZ</i>	Tagatose 6-phosphate aldolase 2	0.774	-0.605	-1.380	Sal	AH
<i>gltA</i>	Citrate synthase	0.288	-0.559	-0.846	PQ, Sal	AH
<i>gltB</i>	Glutamate synthase, large subunit	0.846	-1.425	-2.271	Sal	AH
<i>grxA</i>	Glutaredoxin 1	1.666	-0.106	-1.772	H ₂ O ₂	AH
<i>gshB</i>	Glutathione synthetase	0.768	-0.081	-0.848	Sal	AH
<i>hdeA</i>	Periplasmic acid chaperone	0.841	-0.326	-1.167	Sal	AH
<i>hdeB</i>	Periplasmic acid chaperone	0.782	-0.622	-1.404	Sal	AH
<i>hisF</i>	Cyclase component of IGP synthase	0.150	0.703	0.553	PQ-	BH
<i>ibpB</i>	Chaperone, HSP20 family	-1.691	-2.618	-0.928	H ₂ O ₂	BL
<i>katG</i>	Catalase hydrogen peroxidase 1	-0.578	-0.313	0.265	H ₂ O ₂	AL
<i>lamB</i>	Maltose high-affinity uptake	-4.881	-3.735	1.146	PQ	NH
<i>lldP</i>	L-Lactate permease	1.527	0.219	-1.307	Sal	NL
<i>lysU</i>	Lysyl tRNA synthetase, inducible	0.544	-0.077	-0.621	Sal	AH
<i>malE</i>	Maltose-binding protein, periplasmic	-3.425	-2.933	0.491	PQ	NH
<i>malK</i>	Maltose transport complex, ATP-binding subunit	-4.790	-3.748	1.041	PQ	NH
<i>manX</i>	PTS family, mannose-specific enzyme IIA component	0.000	0.517	0.517	Sal	AL
<i>map</i>	Methionine aminopeptidase	0.510	0.056	-0.454	PQ	AH
<i>marA</i>	Multiple antibiotic resistance	1.321	-0.516	-1.836	PQ, Sal	AH
<i>marB</i>	Regulator for <i>mar</i>	0.894	-0.172	-1.066	Sal	AH
<i>marR</i>	Repressor of <i>mar</i>	0.352	-0.437	-0.789	Sal	AH
<i>mdaB</i>	Drug activity modulator	0.444	-0.223	-0.667	Sal	AH
<i>murF</i>	D-Alanyl:D-alanine adding to cell wall	0.216	-0.170	-0.386	PQ	AH
<i>nfnB</i>	Nitrofurantoin resistance; nitroreductase	0.818	-0.204	-1.022	PQ, Sal	AH
<i>nuoI</i>	NADH dehydrogenase I subunit	0.792	0.310	-0.481	PQ	NL
<i>nuoK</i>	NADH dehydrogenase I subunit	0.766	0.460	-0.306	PQ	NL
<i>ompF</i>	Outer membrane porin	-2.834	-0.892	1.942	PQ-	AL
<i>ompT</i>	Outer membrane protease VII	-0.815	-1.204	-0.389	Sal-	BL
<i>pdhR</i>	Repressor of <i>pdh</i>	0.755	-0.109	-0.863	PQ	AH
<i>pepN</i>	Aminopeptidase N	0.671	0.118	-0.553	Sal	NL
<i>pfkB</i>	Pyruvate formate lyase I (anaerobic)	0.812	0.578	-0.234	Sal	NL
<i>pgi</i>	Glucose phosphate isomerase	0.654	0.177	-0.477	PQ	NL
<i>ptsG</i>	PTS family IIC, glucose specific	0.841	0.554	-0.287	PQ	NL
<i>putA</i>	Proline dehydrogenase	-1.062	-0.353	0.708	Sal	AL
<i>pyrB</i>	Aspartate transcarbamylase	0.133	0.608	0.475	PQ-, Sal-	BH
<i>sdhB</i>	Succinate dehydrogenase	1.833	0.329	-1.505	PQ	AH
<i>tnaA</i>	Tryptophanase	-3.805	0.223	4.028	H ₂ O ₂	AL
<i>treB</i>	Tre-specific PTS enzyme II	-3.156	0.739	3.895	Sal-	AL
<i>yahA</i>	Putative repressor	-0.667	0.422	1.089	PQ-	AL
<i>yaiA</i>	Function unknown	1.154	0.416	-0.738	H ₂ O ₂	NL
<i>ybjC</i>	Function unknown	0.463	-0.447	-0.910	PQ, Sal	AH
<i>ycfR</i>	Function unknown	0.537	-0.819	-1.356	H ₂ O ₂	AH
<i>yfiA</i>	Stabilizes ribosome against dissociation	-1.782	0.581	2.364	PQ-, Sal, H ₂ O ₂	AL
<i>yggJ</i>	Function unknown	0.576	-0.178	-0.755	Sal	AH
<i>yqjD</i>	Function unknown	-0.347	0.378	0.725	Sal	AL
<i>yncE</i>	Function unknown	-0.476	0.331	0.808	PQ, Sal	AL

^a Oxidative response is based on data in references 54 and 84. Induction was by H₂O₂, paraquat (PQ), or sodium salicylate (Sal). Repression is indicated by minus sign (Sal-, PQ-).

^b Values in boldface indicate significance ($\alpha = 0.001$).

^c AH, Acid High; AL, Acid Low; BH, Base High; BL, Base Low; NH, Neutral High; NL, Neutral Low.

^d GAPDH, glyceraldehyde-3-phosphate dehydrogenase.

^e PTS, phosphotransferase.

TABLE 6. Envelope and periplasmic genes

Gene	Function	Log ₂ pH ratio ^a			Class ^b
		5/7	8.7/7	8.7/5	
<i>artI</i>	Periplasmic binding protein of Arg transport system	0.286	0.655	0.369	BH
<i>artM</i>	Arginine periplasmic binding protein	-0.115	0.293	0.407	BH
<i>btuB</i>	B-12 transporter, outer membrane receptor	-0.667	0.037	0.704	AL
<i>cirA</i>	Colicin I receptor production	-0.885	0.260	1.145	AL
<i>clpX</i>	ATPase subunit of ClpXP protease	-0.361	-0.625	-0.264	BL
<i>cpxA</i>	Periplasmic stress sensor (CpxAR)	-0.461	0.230	0.691	AL
<i>cpxP</i>	CpxAR-regulated periplasmic stress protein	-3.232	1.002	4.234	AL
<i>cpxR</i>	Periplasmic stress response regulator (CpxAR)	-0.662	0.590	1.251	AL
<i>dsbA</i>	Thiol:disulfide interchange, periplasmic	0.086	1.117	1.031	BH
<i>dsbC</i>	Disulfide bond isomerase, periplasmic chaperone	-0.288	-0.534	-0.246	BL
<i>fadL</i>	Fatty acid transport, outer membrane	-0.861	1.069	1.931	BH
<i>fecA</i>	Outer membrane ferric citrate receptor	-2.608	-1.128	1.480	NH
<i>fecB</i>	Periplasmic ferric citrate-binding protein	-2.842	-1.538	1.304	NH
<i>fehA</i>	Ferrienterobactin outer membrane receptor	-0.966	-0.289	0.676	NH
<i>fhuD</i>	Ferric hydroxamate binding protein; hydroxamate-dependent iron uptake	-0.767	-0.180	0.587	AL
<i>fliY</i>	Cystine-binding protein, periplasmic	0.233	-0.252	-0.484	AH
<i>hdeA</i>	Acid periplasmic chaperone	0.841	-0.326	-1.167	AH
<i>hdeB</i>	Acid periplasmic protein	0.782	-0.622	-1.404	AH
<i>hisJ</i>	High-affinity histidine-binding protein	-0.149	0.400	0.549	BH
<i>hlpA</i>	Periplasmic chaperone for OMPs ^c	0.099	-0.661	-0.759	BL
<i>lamB</i>	Maltoporin, maltose high-affinity uptake; phage lambda receptor	-4.881	-3.735	1.146	NH
<i>lon</i>	DNA-binding, ATP-dependent protease	-0.600	-1.666	-1.065	BL
<i>malE</i>	Maltose-binding protein, periplasmic	-3.425	-2.933	0.491	NH
<i>malM</i>	Maltose operon periplasmic protein	-4.643	-3.780	0.863	NH
<i>mltB</i>	Membrane-bound murein hydrolase	-0.739	-0.419	0.320	NH
<i>nmpC</i>	Outer membrane	-4.012	-0.051	3.961	AL
<i>ompF</i>	Outer membrane porin protein 1a	-2.834	-0.892	1.942	AL
<i>ompT</i>	Outer membrane protease VII	-0.815	-1.204	-0.389	BL
<i>ompX</i>	OMP, induces RNAP-sigma E	1.523	0.439	-1.083	NL
<i>oppA</i>	Periplasmic oligopeptide binding protein	1.358	0.406	-0.953	NL
<i>pal</i>	Lipoprotein associated with peptidoglycan	0.532	0.379	-0.153	NL
<i>potD</i>	Spermidine-binding membrane protein; regulates <i>pot</i>	0.053	0.509	0.456	BH
<i>ppiA</i>	Rotamase; peptidylprolyl- <i>cis-trans</i> -isomerase A	-0.526	0.231	0.757	AL
<i>pstS</i>	High-affinity, periplasmic phosphate binding protein	-2.479	-2.262	0.217	NH
<i>rbsB</i>	D-Ribose binding protein, periplasmic	-0.749	-1.150	-0.401	BL
<i>rseB</i>	Periplasmic, binds RseA; enhances RpoE-RseA cytoplasmic complex formation	0.631	-0.105	-0.736	AH
<i>secD</i>	SecDF-YajC inner membrane secretion complex	-0.165	0.287	0.452	AL
<i>surA</i>	Periplasmic outer membrane porin chaperone, stationary phase	0.310	0.209	-0.101	NL
<i>tatA</i>	Twin arginine translocation	0.043	0.679	-0.636	BH
<i>tatB</i>	Twin arginine translocation	0.075	0.446	0.372	BH
<i>tolB</i>	Group A colicin uptake and tolerance	0.498	0.421	-0.077	NL
<i>tpX</i>	Thiol peroxidase, antioxidant	0.637	0.178	-0.459	NL
<i>tsx</i>	Phage T6, colicin K resistance; nucleoside channel	-0.707	-0.904	-0.196	BL
<i>ybgF</i>	TolA-binding periplasmic protein	0.312	-0.007	-0.318	AH
<i>ycel</i>	Function unknown; periplasmic protein	-0.036	1.038	1.074	BH
<i>yhcN</i>	Periplasmic protein	3.064	-1.136	-4.199	AH

^a Values in boldface indicate significance ($\alpha = 0.001$).

^b AH, Acid High; AL, Acid Low; BH, Base High; BL, Base Low; NH, Neutral High; NL, Neutral Low.

^c OMPs, outer membrane proteins.

^d RNAP, RNA polymerase.

(4, 68), as are deaminases such as *sdaA* (82). In preliminary experiments, we have repeated our microarray study on cultures grown anaerobically. Under anaerobiosis, several amino acid decarboxylases and deaminases show pH-dependent expression (Hayes and Slonczewski, unpublished).

Stress responses. Several stress responses are known to interact with pH stress and pH resistance, including oxidative stress, heat shock, and envelope stress (for reviews see references 21 and 68). The overlap with salicylate stress could be explained in part by salicylate's effect as a permeant acid, stressing internal pH (60). The *mar* drug resistance operon is known to be coinduced by aromatic permeant acids and low pH (69) under regulation by MarR as well as by the superoxide regulator SoxRA (57).

Beyond salicylate, however, a large number of oxidative stress genes inducible by H₂O₂ or by paraquat showed significant pH-dependent expression, nearly all induced by acid or repressed by base. This finding confirms our hypothesis of a strong connection between acid stress and oxidative stress. Since so much of aerobic respiration is stepped up at pH 5, including cytochrome *o* oxidase, it is likely that acid conditions accelerate the production of oxygen radicals, thus inducing a partial oxidative stress response.

Various envelope and periplasmic stress responses are induced by acid, contributing to acid resistance; the best characterized in terms of mechanism is the acid-induced periplasmic chaperone HdeA (23). Extracellular acid induces a dimer-to-monomer transition in HdeA, which then suppresses aggrega-

TABLE 7. Universal stress and heat shock response genes

Gene	Function	Log ₂ pH ratio ^a			Class ^b
		5/7	8.7/7	8.7/5	
<i>ahpC</i>	Alkyl hydroperoxide reductase	1.003	0.436	-0.568	NL
<i>ahpF</i>	NAD(P)H:peroxiredoxin oxidoreductase	0.777	-0.222	-0.999	AH
<i>cfa</i>	Cyclopropane fatty acid synthase; acid resistance in stationary phase	2.075	-0.480	-2.555	AH
<i>clpB</i>	ClpB protease, ATP-dependent chaperone	-0.219	-1.963	-1.744	BL
<i>cysK</i>	Cysteine synthase, <i>o</i> -acetylserine sulfhydrylase A	1.204	1.351	0.147	NL
<i>cysZ</i>	Unknown function	0.440	0.212	-0.228	NL
<i>dinI</i>	Inhibits RecA coprotease	0.132	0.568	0.436	BH
<i>dinJ</i>	Induced by DNA damage	0.683	1.192	0.508	BH
<i>dnaJ</i>	DnaK cochaperone	-0.908	-2.155	-1.247	BL
<i>dnaK</i>	HSP-70-type molecular chaperone	-0.894	-2.249	-1.356	BL
<i>dps</i>	Stress response DNA-binding protein	1.130	0.105	-1.025	NL
<i>grpE</i>	Nucleotide exchange factor for DnaKJ	-0.491	-1.141	-0.650	BL
<i>grxA</i>	Glutaredoxin 1	1.666	-0.106	-1.772	AH
<i>hdeA</i>	Acid periplasmic chaperone	0.841	-0.326	-1.167	AH
<i>hdeB</i>	Acid periplasmic chaperone	0.782	-0.622	-1.404	AH
<i>hslI</i>	Heat-inducible novobiocin resistance	0.368	-0.676	-1.044	AH
<i>hslU</i>	Heat-inducible ATP-dependent protease	-0.745	-1.688	-0.946	BL
<i>hslV</i>	Heat-inducible ATP-dependent protease	-1.125	-1.913	-0.788	BL
<i>ibpB</i>	Heat-inducible chaperone, HSP20 family	-1.691	-2.618	-0.928	BL
<i>hslO</i>	Hsp33, cytoplasmic heat shock chaperone activated by disulfide bond formation	-1.453	-1.737	-0.284	BL
<i>hslR (yrfH)</i>	Hsp15, heat shock, binds RNA and DNA	-1.773	-1.940	-0.168	NH
<i>katG</i>	Catalase-hydrogen peroxidase I	-0.578	-0.313	0.265	AL
<i>rpoE</i>	RNAP ^c sigma E, envelope heat stress	0.310	-0.427	-0.738	AH
<i>rpoH</i>	RNAP sigma 32, heat shock regulons	-0.378	0.084	0.462	AL
<i>rseA</i>	Anti-RpoE sigma factor, spans inner membrane	0.419	-0.159	-0.578	AH
<i>rseB</i>	Binds periplasmic domain of anti-RpoE sigma RseA	0.631	-0.105	-0.736	AH
<i>sodB</i>	Superoxide dismutase, Fe; acid inducible	0.752	0.280	-0.472	NL
<i>yedB</i>	Function unknown, peroxidase homolog	0.654	-0.329	-0.983	AH
<i>yedO</i>	Acid inducible, function unknown	0.842	-0.625	-1.467	AH
<i>uspD (yiiT)</i>	UV resistance	1.333	-0.497	-1.830	AH

^a Values in boldface indicate significance ($\alpha = 0.001$).

^b AH, Acid High; AL, Acid Low; BH, Base High; BL, Base Low; NH, Neutral High; NL, Neutral Low.

^c RNAP, RNA polymerase.

tion by acid-denatured proteins. Our study reveals additional potential contributors to acid resistance and base resistance, including genes of unknown function such as *yhcN*, induced by acid, and *yceI*, induced by base.

Our study presents the most comprehensive picture to date of acid and base response by *E. coli* grown aerobically in complex medium. Overall, low pH accelerates acid consumption and proton export, while coinducing oxidative stress, possibly through increased production of oxygen radicals. High pH accelerates proton import while repressing the energy-expensive systems of flagellar biosynthesis and chemotaxis. Finally, pH differentially regulates a large number of periplasmic and envelope stress systems, as well as transporters, chaperones, and redox regulators.

ACKNOWLEDGMENTS

The class comparison and cluster analysis were performed using BRB ArrayTools v3.1 developed by Richard Simon and Amy Peng Lam. We thank Bryan Lin and Ariel Kahrl for excellent technical assistance.

This work was supported by grant MCB-0234732 from the National Science Foundation and by undergraduate research funds from the Kenyon College grant from the Howard Hughes Medical Institute Biological Sciences Education Program.

REFERENCES

- Ades, S. E., I. L. Grigорова, and C. A. Gross. 2003. Regulation of the alternative sigma factor σ^E during initiation, adaptation, and shutoff of the extracytoplasmic heat shock response in *Escherichia coli*. *J. Bacteriol.* **185**: 2512–2519.
- Argaman, L., R. Hershberg, J. Vogel, G. Bejerano, E. Gerhart, H. Wagner, H. Margalit, and S. Altuvia. 2001. Novel small RNA-encoding genes in the intergenic regions of *Escherichia coli*. *Curr. Biol.* **11**:941–950.
- Arnold, C. N., J. McElhanon, A. Lee, R. Leonhart, and D. A. Siegle. 2001. Global analysis of *Escherichia coli* gene expression during the acetate-induced acid tolerance response. *J. Bacteriol.* **183**:2178–2186.
- Auger, E. A., K. E. Redding, T. Plumb, L. C. Childs, S. Y. Meng, and G. N. Bennett. 1989. Construction of *lac* fusions to the inducible arginine and lysine decarboxylase genes of *Escherichia coli* K-12. *Mol. Microbiol.* **3**:609–620.
- Bakker, E. P., and W. E. Mangerich. 1981. Interconversion of components of the bacterial proton motive force by electrogenic potassium transport. *J. Bacteriol.* **147**:820–826.
- Bearson, B. L., L. Wilson, and J. W. Foster. 1998. A low pH-inducible, PhoPQ-dependent acid tolerance response protects *Salmonella typhimurium* against inorganic acid stress. *J. Bacteriol.* **180**:2409–2417.
- Behari, J., L. Stagon, and S. B. Calderwood. 2001. *pepA*, a gene mediating pH regulation of virulence genes in *Vibrio cholerae*. *J. Bacteriol.* **183**:178–188.
- Bingham, R. J., K. S. Hall, and J. L. Slonczewski. 1990. Alkaline induction of a novel gene locus, *alk*, in *Escherichia coli*. *J. Bacteriol.* **172**:2184–2186.
- Blankenhorn, D., J. Phillips, and J. L. Slonczewski. 1999. Acid- and base-induced proteins during aerobic and anaerobic growth of *Escherichia coli* revealed by two-dimensional gel electrophoresis. *J. Bacteriol.* **181**:2209–2216.
- Buurman, E. T., D. McLaggan, J. Naprstek, and W. Epstein. 2004. Multiple paths for nonphysiological transport of K⁺ in *Escherichia coli*. *J. Bacteriol.* **186**:4238–4245.
- Castanie-Cornet, M., T. A. Penfound, D. Smith, J. Elliott, and J. W. Foster. 1999. Control of acid resistance in *Escherichia coli*. *J. Bacteriol.* **181**:3525–3535.
- Castanie-Cornet, M., and J. W. Foster. 2001. *Escherichia coli* acid resistance: cAMP receptor protein and a 20 bp cis-acting sequence control pH and stationary phase expression of the *gadA* and *gadBC* glutamate decarboxylase genes. *Microbiology* **147**:709–715.
- Chagneau, C., M. Heyde, S. Alonso, R. Portulier, and P. Laloi. 2001. External-pH-dependent expression of the maltose regulon and *ompF* gene in

- Escherichia coli* is affected by the level of glycerol kinase, encoded by *glpK*. J. Bacteriol. **183**:5675–5683.
14. Cotter, P. A., V. Chepuri, R. B. Gennis, and R. P. Gunsalus. 1990. Cytochrome *o* (*cyoABCDE*) and *d* (*cydAB*) oxidase gene expression in *Escherichia coli* is regulated by oxygen, pH, and the *fnr* gene product. J. Bacteriol. **172**:6333–6338.
 15. Cummings, J. H., E. W. Pomare, W. J. Branch, C. P. Naylor, and G. T. Macfarlane. 1997. Short chain fatty acids in human large intestine, portal, hepatic and venous blood. Gut **28**:1221–1227.
 16. DiGiuseppe, P. A., and T. J. Silhavy. 2003. Signal detection and target gene induction by the CpxRA two-component system. J. Bacteriol. **185**:2432–2440.
 17. Drasar, B. S., M. Shiner, and G. M. McLeod. 1969. Studies on the intestinal flora. I. The bacterial flora of the gastrointestinal tract in healthy and achlorhydric persons. Gastroenterology **56**:71–79.
 18. Dressman, J. B., R. R. Berardi, L. C. Dermentzoglou, T. L. Russel, S. P. Schmaltz, J. L. Barnett, and K. M. Jarvenpaa. 1990. Upper gastrointestinal (GI) pH in young, healthy men and women. Pharm. Res. **7**:756–761.
 19. Eisen, M. B., P. T. Spellman, P. O. Brown, and D. Botstein. 1998. Cluster analysis and display of genome-wide expression patterns. Proc. Natl. Acad. Sci. USA **95**:14863–14868.
 20. Evans, D. F., G. Pye, R. Bramley, A. G. Clark, T. J. Dyson, and J. D. Hardcastle. 1988. Measurement of gastrointestinal pH profiles in normal ambulant human subjects. Gut **29**:1035–1041.
 21. Foster, J. W. 2000. Microbial responses to acid stress, p. 99–115. In G. Storz and R. Hengge-Aronis (ed.), Bacterial stress responses. ASM Press, Washington, D.C.
 22. Francez-Charlot, A., B. Laugel, A. Van Gemert, N. Dubarry, F. Wiorowski, M. P. Castanie-Cornet, C. Gutierrez, and K. Cam. 2003. RcsCDB His-Asp phosphorelay system negatively regulates the *flhDC* operon in *Escherichia coli*. Mol. Microbiol. **49**:823–832.
 23. Gajiwala, K. S., and S. K. Burley. 2000. HDEA, a periplasmic protein that supports acid resistance in pathogenic enteric bacteria. J. Mol. Biol. **295**:605–612.
 24. Gerchman, Y., Y. Olami, A. Rimon, D. Taglicht, and S. Schuldiner. 1993. Histidine-226 is part of the pH sensor of NhaA, a Na⁺/H⁺ antiporter in *Escherichia coli*. Proc. Natl. Acad. Sci. USA **90**:1212–1216.
 25. Giannella, R. A., S. A. Broitman, and N. Zamcheck. 1973. Influence of gastric acidity on bacterial and parasitic enteric infections. A perspective. Ann. Intern. Med. **78**:271–276.
 26. Gong, F., and C. Yanofsky. 2002. Instruction of translating ribosome by nascent peptide. Science **297**:1864–1867.
 27. Gong, S., H. Richard, and J. W. Foster. 2003. YjdE (AdiC) is the arginine: agmatine antiporter essential for arginine-dependent acid resistance in *Escherichia coli*. J. Bacteriol. **185**:4402–4409.
 28. Gross, C. A. 1996. Function and regulation of the heat shock proteins, p. 1382–1399. In F. C. Neidhardt, R. Curtiss III, J. L. Ingraham, E. C. C. Lin, K. B. Low, B. Magasanik, W. S. Reznikoff, M. Riley, M. Schaechter, and H. E. Umbarger (ed.), *Escherichia coli* and *Salmonella*: cellular and molecular biology, 2nd ed., vol. 1. ASM Press, Washington, D.C.
 29. Harold, F. M., and P. C. Maloney. 1996. Energy transduction by ion currents, p. 283–306. In F. C. Neidhardt, R. Curtiss III, J. L. Ingraham, E. C. C. Lin, K. B. Low, B. Magasanik, W. S. Reznikoff, M. Riley, M. Schaechter, and H. E. Umbarger (ed.), *Escherichia coli* and *Salmonella*: cellular and molecular biology, 2nd ed. ASM Press, Washington, D.C.
 30. Hersh, B. M., F. T. Farooq, D. N. Barstad, D. Blankenhorn, and J. L. Slonczewski. 1996. A glutamate-dependent acid resistance gene in *Escherichia coli*. J. Bacteriol. **178**:3978–3981.
 31. Heyde, M., P. Laloi, and R. Portalier. 2000. Involvement of carbon source and acetyl phosphate in the external-pH-dependent expression of porin genes in *Escherichia coli*. J. Bacteriol. **182**:198–202.
 32. Kasimoglu, E., S. Park, J. Malek, C. P. Tseng, and R. P. Gunsalus. Transcriptional regulation of the proton-translocating ATPase (*atpIBEFHAG DC*) operon of *Escherichia coli*: control by cell growth. J. Bacteriol. **178**:5563–5567.
 33. Khan, S., and R. Macnab. 1980. Proton chemical potential, proton electrical potential and bacterial motility. J. Mol. Biol. **138**:599–614.
 34. Kihara, M., and R. Macnab. 1981. Cytoplasmic pH mediates pH taxis and weak-acid repellent taxis of bacteria. J. Bacteriol. **145**:1209–1221.
 35. Kirkpatrick, C., L. M. Maurer, N. E. Oyelakin, Y. N. Yoncheva, R. Maurer, and J. L. Slonczewski. 2001. Acetate and formate stress: opposite responses in the proteome of *Escherichia coli*. J. Bacteriol. **183**:6466–6477.
 36. Kothapalli, R., S. J. Yoder, S. Mane, and T. P. Loughran, Jr. 2002. Microarray results: how accurate are they? BMC Bioinformatics **3**:22.
 37. Kutsukake, K. 1997. Autogenous and global control of the flagellar master operon *flhD*, in *Salmonella typhimurium*. Mol. Gen. Genet. **254**:440–448.
 38. Lambert, L. A., K. Abshire, D. Blankenhorn, and J. L. Slonczewski. 1997. Proteins induced in *Escherichia coli* by benzoic acid. J. Bacteriol. **179**:7595–7599.
 39. Li, C., and W. H. Wong. 2001. Model-based analysis of oligonucleotide arrays: expression index computation and outlier detection. Proc. Natl. Acad. Sci. USA **98**:31–36.
 40. Li, C., and W. H. Wong. 2003. DNA-chip analyzer (dChip), p. 120–141. In G. Parmigiani, E. S. Garrett, R. Irizarry, and S. L. Zeger (ed.), The analysis of gene expression data: methods and software. Springer-Verlag, New York, N.Y.
 41. Ma, Z., H. Richard, D. L. Tucker, T. Conway, and J. W. Foster. 2002. Collaborative regulation of *Escherichia coli* glutamate-dependent acid resistance by two AraC-like regulators, GadX and GadW (YhiW). J. Bacteriol. **184**:7001–7012.
 42. Macnab, R. M. 2003. How bacteria assemble flagella. Annu. Rev. Microbiol. **57**:77–100.
 43. Masse, E., N. Majdalani, and S. Gottesman. 2003. Regulatory roles for small RNAs in bacteria. Curr. Opin. Microbiol. **6**:120–124.
 44. Masuda, N., and G. Church. 2003. Regulatory network of acid resistance in *Escherichia coli*. Mol. Microbiol. **48**:699–712.
 45. McFall, E., and E. B. Newman. 1996. Amino acids as carbon sources, p. 358–379. In F. C. Neidhardt, R. Curtiss III, J. L. Ingraham, E. C. C. Lin, K. B. Low, B. Magasanik, W. S. Reznikoff, M. Riley, M. Schaechter, and H. E. Umbarger (ed.), *Escherichia coli* and *Salmonella*: cellular and molecular biology, 2nd ed. ASM Press, Washington, D.C.
 46. Minamino, T., Y. Imae, F. Oosawa, Y. Kobayashi, and K. Oosawa. 2003. Effect of intracellular pH on rotational speed of bacteria flagellar motors. J. Bacteriol. **185**:1190–1194.
 47. Neely, M. N., C. L. Dell, and E. R. Olson. 1994. Roles of LysP and CadC in mediating the lysine requirement for acid induction of the *Escherichia coli cad* operon. J. Bacteriol. **176**:3278–3285.
 48. Nikaïdo, H. 1996. Outer membrane, p. 29–47. In F. C. Neidhardt, R. Curtiss III, J. L. Ingraham, E. C. C. Lin, K. B. Low, B. Magasanik, W. S. Reznikoff, M. Riley, M. Schaechter, and H. E. Umbarger (ed.), *Escherichia coli* and *Salmonella*: cellular and molecular biology, 2nd ed. ASM Press, Washington, D.C.
 49. Oliver, D. B. 1996. Periplasm, p. 88–103. In F. C. Neidhardt, R. Curtiss III, J. L. Ingraham, E. C. C. Lin, K. B. Low, B. Magasanik, W. S. Reznikoff, M. Riley, M. Schaechter, and H. E. Umbarger (ed.), *Escherichia coli* and *Salmonella*: cellular and molecular biology, 2nd ed. ASM Press, Washington, D.C.
 50. Payne, D., D. Tatham, E. D. Williamson, and R. W. Titball. 1998. The pH 6 antigen of *Yersinia pestis* binds to β 1-linked galactosyl residues in glycosphingolipids. Infect. Immun. **66**:4545–4548.
 51. Piper, M. D. W., P. Daran-Lipujade, C. Bro, B. Regenber, S. Knudsen, J. Nielsens, and J. T. Pronk. 2002. Reproducibility of oligonucleotide microarray transcriptome analyses. J. Biol. Chem. **277**:37001–37008.
 52. Plack, R. H., Jr., and B. P. Rosen. 1980. Cation/proton antiport systems in *Escherichia coli*. Absence of potassium/proton antiporter activity in a pH-sensitive mutant. J. Biol. Chem. **255**:3824–3825.
 53. Polen, T., D. Rittmann, V. F. Wendisch, and H. Sahn. 2003. DNA microarray analysis of the long-term adaptive response of *Escherichia coli* to acetate and propionate. Appl. Environ. Microbiol. **69**:1759–1774.
 54. Pomposiello, P., M. H. J. Bennik, and B. Dimple. 2001. Genome-wide transcriptional profiling of *Escherichia coli* responses to superoxide stress and sodium salicylate. J. Bacteriol. **183**:3890–3920.
 55. Repaske, D., and J. Adler. 1981. Change in intracellular pH of *Escherichia coli* mediates the chemotactic response to certain attractants and repellents. J. Bacteriol. **145**:1196–1208.
 56. Roe, A. J., D. McLaggan, I. Davidson, C. O'Byrne, and I. R. Booth. 1998. Perturbations of anion balance during inhibition of growth of *Escherichia coli* by weak acids. J. Bacteriol. **180**:767–772.
 57. Rosner, J. L., and J. L. Slonczewski. 1994. Dual regulation of *inaA* by the multiple antibiotic resistance (*mar*) and superoxide (*soxRS*) stress response systems of *Escherichia coli*. J. Bacteriol. **176**:6262–6269.
 58. Rowbury, R. J., Z. Lazim, and M. Goodson. 1996. Regulatory aspects of alkali tolerance induction in *Escherichia coli*. Lett. Appl. Microbiol. **22**:429–432.
 59. Russell, J. B., and F. Diez-Gonzalez. 1998. The effect of fermentation acids on bacterial growth. Adv. Microb. Physiol. **39**:205–234.
 60. Salmond, C. V., R. G. Kroll, and I. Booth. 1984. The effect of food preservatives on pH homeostasis in *Escherichia coli*. J. Gen. Microbiol. **130**:2845–2850.
 61. Schadt, E. E., C. Li, B. Ellis, and W. Wong. 2001. Feature extraction and normalization algorithms for high-density oligonucleotide gene expression array data. J. Cell. Biochem. **84**(S37):120–125.
 62. Schadt, E. E., C. Li, C. Su, and W. H. Wong. 2001. Analyzing high-density oligonucleotide gene expression array data. J. Cell. Biochem. **80**:192–202.
 63. Schuldiner, S., V. Agmon, J. Brandsma, A. Cohen, E. Friedman, and E. Padman. 1986. Induction of SOS function by alkaline intracellular pH in *Escherichia coli*. J. Bacteriol. **168**:936–939.
 64. Shi, W., Y. Zhou, J. Adler, and C. A. Gross. 1992. DnaK, DnaJ, and GrpE are required for flagellum synthesis in *Escherichia coli*. J. Bacteriol. **174**:6256–6263.
 65. Skandamis, P. N., and G.-J. E. Nychas. 2000. Development and evaluation of a model predicting the survival of *Escherichia coli* O157:H7 NCTC 12900 in homemade eggplant salad at various temperatures, pHs, and oregano essential oil concentrations. Appl. Environ. Microbiol. **66**:1646–1653.

66. Slonczewski, J. L., T. N. Gonzalez, F. M. Bartholomew, and N. J. Holt. 1987. Mu d-directed *lacZ* fusions regulated by low pH in *Escherichia coli*. *J. Bacteriol.* **169**:3001-3006.
67. Slonczewski, J. L., R. M. Macnab, J. R. Alger, and A. M. Castle. 1982. Effects of pH and repellent tactic stimuli on protein methylation levels in *Escherichia coli*. *J. Bacteriol.* **152**:384-399.
68. Slonczewski, J. L., and J. W. Foster. 1996. pH-related genes and survival at extreme pH, p. 1539-1552. In F. C. Neidhardt, R. Curtiss III, J. L. Ingraham, E. C. C. Lin, K. B. Low, B. Magasanik, W. S. Reznikoff, M. Riley, M. Schaechter, and H. E. Umbarger (ed.), *Escherichia coli* and *Salmonella*: cellular and molecular biology, 2nd ed. ASM Press, Washington, D.C.
69. Slonczewski, J. L., B. P. Rosen, J. R. Alger, and R. M. Macnab. 1981. pH homeostasis in *Escherichia coli*: measurement by ^{31}P nuclear magnetic resonance of methylphosphonate and phosphate. *Proc. Natl. Acad. Sci. USA* **78**:6271-6275.
70. Small, P., D. Blankenhorn, D. Welty, E. Zinser, and J. L. Slonczewski. 1994. Acid and base resistance in *Escherichia coli* and *Shigella flexneri*: role of *rpoS* and growth pH. *J. Bacteriol.* **176**:1729-1737.
71. Soksawatmaekhin, W., A. Kuraishi, K. Skata, K. Kashiwagi, and K. Igarahi. 2004. Excretion and uptake of cadaverine by CadB and its physiological functions in *Escherichia coli*. *Mol. Microbiol.* **51**:1401-1412.
72. Soutourina, O. A., E. Krin, C. Laurent-Winter, F. Hommais, A. Danchin, and P. N. Bertin. 2002. Regulation of bacterial motility in response to low pH in *Escherichia coli*: the role of H-NS protein. *Microbiology* **148**:1543-1551.
73. Stancik, L. M., D. M. Stancik, B. Schmidt, D. M. Barnhart, Y. N. Yoncheva, and J. L. Slonczewski. 2002. pH-dependent expression of periplasmic proteins and amino acid catabolism in *Escherichia coli*. *J. Bacteriol.* **184**:4246-4258.
74. Taglicht, D., E. Padan, A. B. Oppenheim, and S. Schuldiner. 1987. An alkaline shift induces the heat shock response in *Escherichia coli*. *J. Bacteriol.* **169**:885-887.
75. Taglicht, D., E. Padan, and S. Schuldiner. 1991. Overproduction and purification of a functional Na^+/H^+ antiporter coded by *nhaA* (*ant*) from *Escherichia coli*. *J. Biol. Chem.* **266**:11289-11294.
76. Tomoyasu, T., A. Takaya, E. Isogai, and T. Yamamoto. 2003. Turnover of FlhD and FlhC, master regulator proteins for *Salmonella* flagellum biogenesis, by the ATP-dependent ClpXP protease. *Mol. Microbiol.* **48**:443-452.
77. Tramonti, A., M. Visca, M. De Canio, M. Falconi, and D. De Biase. 2002. Functional characterization and regulation of *gadX*, a gene encoding an AraC/XylS-like transcriptional activator of the *Escherichia coli* glutamic acid decarboxylase system. *J. Bacteriol.* **184**:2603-2613.
78. Tucker, D. L., N. Tucker, and T. Conway. 2002. Gene expression profiling of the pH responses in *Escherichia coli*. *J. Bacteriol.* **184**:6551-6558.
79. Tucker, D. L., N. Tucker, Z. Ma, J. W. Foster, R. L. Miranda, P. S. Cohen, and T. Conway. 2003. Genes of the GadX-GadW regulon in *Escherichia coli*. *J. Bacteriol.* **185**:3190-3201.
80. White, S., F. E. Tuttle, D. Blankenhorn, D. C. Dosch, and J. L. Slonczewski. 1992. pH dependence and gene structure of *inaA* in *Escherichia coli*. *J. Bacteriol.* **174**:1537-1543.
81. Yeung, K. Y., and W. L. Ruzzo. 2001. Principal component analysis for clustering gene expression data. *Bioinformatics* **17**:763-774.
82. Yohannes, E., D. M. Barnhart, and J. L. Slonczewski. 2004. pH-dependent catabolic protein expression during anaerobic growth of *Escherichia coli* K-12. *J. Bacteriol.* **186**:192-199.
83. Yuen, T., E. Wurmbach, R. L. Pfeffer, B. J. Ebersole, and S. C. Sealfon. 2002. Accuracy and calibration of commercial oligonucleotide and custom cDNA microarrays. *Nucleic Acids Res.* **30**:1-9.
84. Zheng, M., X. Wang, L. J. Templeton, D. R. Smulski, R. A. LaRossa, and G. Storz. 2001. DNA microarray-mediated transcriptional profiling of the *Escherichia coli* response to hydrogen peroxide. *J. Bacteriol.* **183**:4562-4570.